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Chemistry

Reviewing the effect of metal complexation upon the antioxidant/antiradical properties of L-ascorbic acid

Revisando o efeito da complexação de metais sobre as propriedades antioxidantes/antirradicais do ácido L-ascórbico

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ABSTRACT

L-ascorbic acid is a molecule used in the hydroxylation of various biochemical reactions in cells. Its main function is the hydroxylation of collagen, the fibrillar protein that gives resistance to bones, teeth, tendons and walls of blood vessels. Furthermore, it is a powerful antioxidant, being used to transform reactive oxygen species into inert forms. It is also used in the synthesis of some molecules that serve as hormones or neurotransmitters. In this review, a series of reactions are presented and discussed with the aim to discuss as some chemical parameters such as pH, redox potential, presence of different metal ions and ascorbic acid works effectively as a ligand. Several mechanisms are revisited and aspects as the effect of transition metals over the redox chemistry of acid is presented.

Keywords: L-ascorbic acid; Redox potential; Metal ligand

RESUMO

O ácido L-ascórbico é uma molécula utilizada na hidroxilação de várias reações bioquímicas nas células. Sua principal função é a hidroxilação do colágeno, a proteína fibrilar que dá resistência aos ossos, dentes, tendões e paredes dos vasos sanguíneos. Além disso, é um poderoso antioxidante, sendo utilizado para transformar espécies reativas de oxigênio em formas inertes. Também é usado na síntese de algumas moléculas que servem como hormônios ou neurotransmissores. Nesta revisão, uma série de reações são apresentadas e discutidas com o objetivo de discutir alguns parâmetros químicos como pH, potencial redox, presença de diferentes íons metálicos e o ácido ascórbico funciona efetivamente como um ligante. Vários mecanismos são revisitados e aspectos como o efeito dos metais de transição sobre a química redox do ácido são apresentados.

Palavras-chave: Ácido L-ascórbico; Potencial redox; Ligante metálico



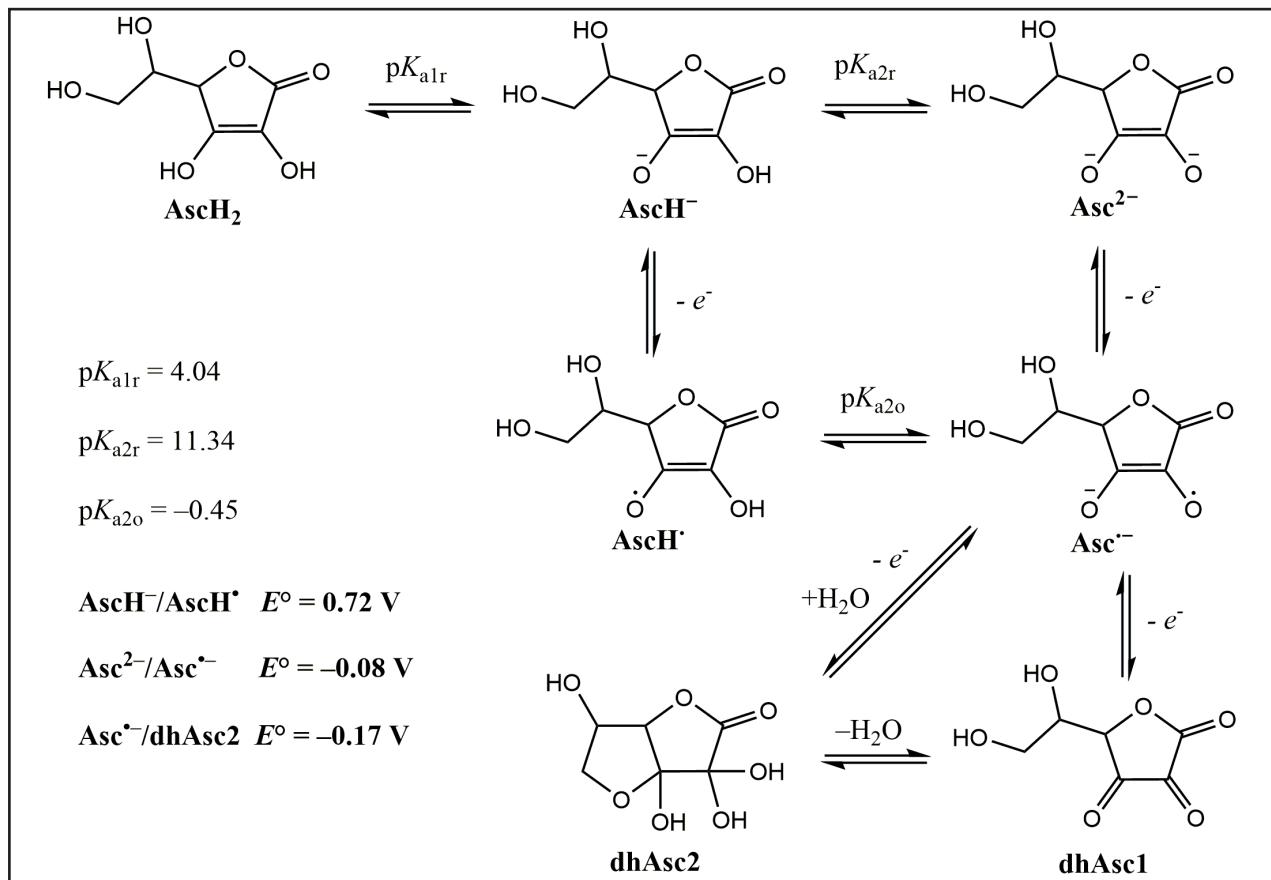
1 INTRODUCTION

L-ascorbic acid (AscH_2) is among the most widely cited forms of water-soluble biological antioxidants. The ability to scavenge free radicals appears, in part, to involve one-electron oxidation where ascorbate serves as reductant towards radical species (Crans *et al.*, 2008). Reaction of AscH_2 with peroxy or hydroxyl radicals typically yields radical intermediate species, which can be subsequently quenched as part of an overall antioxidant effect (Yin *et al.*, 2022). On the basis of the redox potential of AscH_2 , radical intermediates seem to be formed with nearly equal facility during radical scavenging reactions (Tu, Njus and Schlegel, 2017). Nevertheless, AscH_2 is also known to act as a pro-oxidant in fats, particularly in aqueous fat systems (Kanner, Mendel and Budowski, 1977).

Metal ions appear to be involved in the pro-oxidant activity of AscH_2 , as shown by the inhibition of such effect by metal chelating compounds as ethylenediaminetetraacetic acid (EDTA) or polyphosphates (Timoshnikov *et al.*, 2022). Indeed, Fe^{3+} and Cu^{2+} have been reported to accelerate the pro-oxidant activity of AscH_2 towards lipids (Ritacca *et al.*, 2022).

The first standard reduction potential (E°) of AscH_2 is around 0.72 V, while the second one is around -0.17 V; this means that, even though it is a great anti-radical agent, it is not necessarily a good reducing agent. Free radicals are unstable and have a high E° , allowing electron transfer (ET) from AscH_2 . Its antioxidant character is linked to the availability of electrons to reduce strong oxidizing agents, as free radicals, but it is not intended to behave as a strong reducing agent in the biological environment (Tu, Njus and Schlegel, 2017).

The antioxidant potential of AscH_2 comes from its ability to be converted into dehydroascorbate (dhAsc) through the abstraction of two protons and two electrons (Figure 1). This redox reaction provides the electrons needed to stabilize radicals, as well as the proton to balance charges (Nimse and Pal, 2015; Pehlivan, 2017).

Figure 1 – Possible mechanisms for the oxidation of AscH_2 to dhAsc

Source: Authors

It is very interesting to note that AscH_2 is an extremely efficient molecule which is adapted to function in the physiological environment. Figure 1 shows that, at a pH close to 7.0, AscH_2 appears almost entirely in its monoprotic AscH^- form ($pK_{\text{a}1\text{r}} = 4.04$), while the $pK_{\text{a}2\text{r}}$ value is 11.34. If AscH^- ($\text{AscH}^-/\text{AscH}^\cdot \quad E^\circ = 0.72 \text{ V}$) meets a strong enough oxidizing agent, it undergoes oxidation by transferring one electron, which results in AscH^\cdot formation.

Once in radical form, it becomes a strong base, promptly losing the second proton ($pK_{\text{a}2\text{o}} = -0.45$ against $pK_{\text{a}2\text{r}} = 11.34$). The $\text{Asc}^{\cdot-}/\text{dhAsc}$ pair is a much more efficient reducer ($E^\circ = -0.174 \text{ V}$), easily transferring the second electron to the substrate.

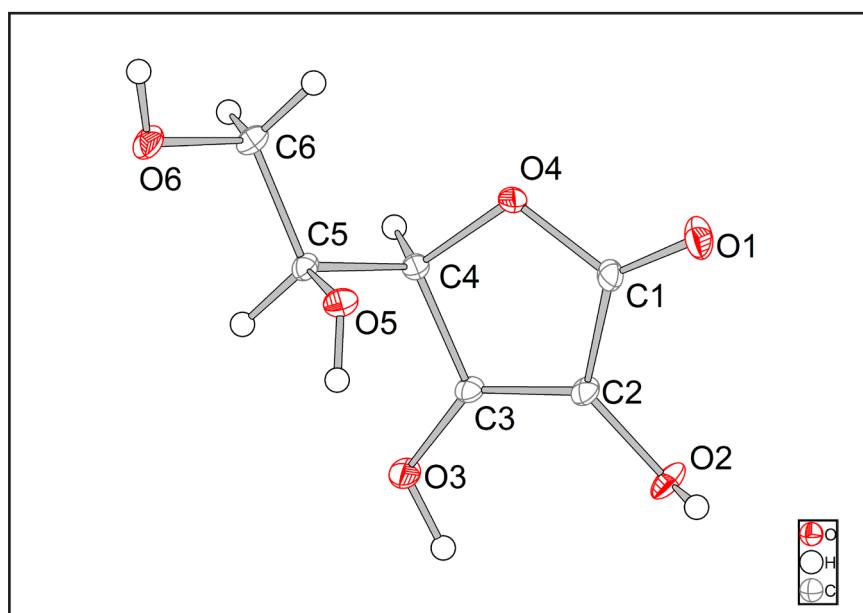
When deprotonated, AscH_2 functions as an O-donor base with low polarizability, a hard base according to the hard and soft acids and bases (HSAB) theory; it easily forms complexes with various metal ions present in solution, mainly transition metals

showing high oxidation states (e.g. Fe^{3+} , Co^{3+} , Cu^{2+} , Cr^{6+} , V^{5+}). Complexation occurs because the AscH^- and Asc^{2-} species must donate electronic density to the metal ions, forming a stable complex. Interaction with the metallic center totally changes the expected redox behavior of AscH_2 .

2 L-ASCORBIC ACID AS A LIGAND

In the nearly 90 years since its discovery (1928), AscH_2 has become “the most famous yet least understood of the vitamins”. The crystal structure of AscH_2 (Figure 2) has been reported by Hvoslef in 1964 (Hvoslef 1964). Despite being a simple molecule, its biochemistry is poorly understood owing to a quite complicated redox chemistry, which makes it both an interesting and intriguing reducing agent in inorganic systems. Many solution studies have since been carried out on reactions between AscH_2 and metal ions. The important work of Martell has established the catalytic role of metals in AscH_2 oxidation (Martell, 1982).

Figure 2 – Crystal structure of L-ascorbic acid



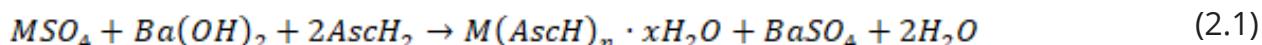
Source: Martell, 1982

As a weak diprotic acid ($pK_{a1} = 4.25$ and $pK_{a2} = 11.79$), the monoanion (AscH^-) is formed at pH 4–5 with deprotonation of O(3)-H, and the dianion (Asc^{2-}) is formed at pH 11–12 with deprotonation of the O(2)-H. The mono-anionic form is more stable due to the delocalization of the negative charge between the oxygen atoms at positions 1 and 3. Although AscH_2 has numerous donor atoms to promote the formation of a metal complex, interaction of AscH^- with metals mainly occurs monodentately through the O(3) atom, or by chelation via O(3) and O(2), depending on the nature of the metal cation and the solution pH. Multiple other bonding modes have been proposed in the solid state, including the participation of the carbonyl oxygen and side chain OH groups.

Stability of complexes is generally less than might have been expected. The formation constants of the 1:1 complexes are in the range of 10 to $10^{3.6}$ (Martell, 1982). The values are quite small, possibly as a result of the low negative charge on the ligand anion. Jabs and Gaube determined the ligand field parameters of the AscH^- ligand and suggested that ascorbate should take an intermediate position in the spectroscopic series, around H_2O and O^{2-} and just before the fluoride ligand in the nephelauxetic series (Jabs and Gaube, 1986). Later, Cieslak-Golonka and co-workers calculated crystal field parameters of some chromium ascorbate complexes for octahedral and tetragonal symmetries from diffuse reflectance spectra (Adach, Janyst and Cieślak-Golonka, 1995). They found that the Dq values are in the $1600\text{--}1800\text{ cm}^{-1}$ region and larger in solution, typical of oxygen ligands.

Nearly all the work on transition metal pure ascorbate complexes has been performed with the first-row metals and on powdered samples. Since no single crystal data is available, the structural assignments have generally been deduced from UV-vis, NMR, IR and magnetic measurements. Due to the unstable nature of the molecule and hydrolytic instabilities of the complexes, there have not been many reports on the isolation of solid complexes of AscH_2 . The proposed structures of the pure ascorbate complexes have been the subject of the most controversy in the absence of X-ray crystal data.

The first systematic synthesis and isolation of binary ascorbate complexes with redox-inert transition metals have been described by Jabs and Gaube (JABS and GAUBE, 1984). Complexes of the type $M(AscH)_n \cdot xH_2O$ ($M = TiO^{2+}$, Cr^{3+} , Mn^{2+} , Co^{2+} , Ni^{2+} and Zn^{2+}) were obtained through the reaction:



Since then, complexes with highly oxidized transition metals have been investigated by other authors. In these cases, the metal center is generally reduced by $AscH_2$ via an inner-sphere reaction. Ferrer and co-workers demonstrated that the primary complexes generated by the interaction of dhAsc with metal ions are not stable and irreversibly hydrolyze to diketogulonic acid complexes of the related metal (Ferrer, Williams and Baran, 1998).

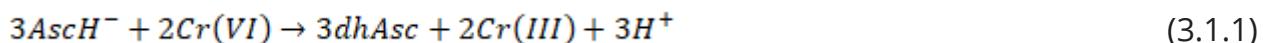
3 EFFECT OF TRANSITION METALS OVER THE REDOX CHEMISTRY OF L-ASCORBIC ACID

Complexation of $AscH_2$ with metal ions having E° greater than 0.72 V, e.g., Co^{3+} , Cr^{6+} and V^{5+} (Table 1), promotes ET from $AscH_2$ to the metal, leading to dhAsc formation. Usually, dhAsc is irreversibly hydrolyzed to diketogulonic acid (Figure 1) (Ferrer, Williams and Baran, 1998; Kontogiorges *et al.*, 2020). In this scenario, there is a reduction in the concentration of $AscH_2$ in the medium, leading to an underestimated measurement of the antioxidant activity, for example. Some of the known redox complexation reactions of $AscH_2$ are:

3.1 High oxidation state metals: Co(III), Cr(VI) and V(V)

Complexation of $AscH_2$ with metal ions having E° greater than 0.72 V, e.g., Co^{3+} , Cr^{6+} and V^{5+} (Table 1), promotes ET from $AscH_2$ to the metal, leading to dhAsc formation. Usually, dhAsc is irreversibly hydrolyzed to diketogulonic acid (Figure 1) (Ferrer, Williams and Baran, 1998;

Kontoghiorghe *et al.*, 2020). In this scenario, there is a reduction in the concentration of Asch₂ in the medium, leading to an underestimated measurement of the antioxidant activity, for example. Some of the known redox complexation reactions of Asch₂ are:



3.2 Redox inert transition metals: Zn(II), Cd(II), and Al(III)

Interaction of these metals with Asch₂ has been assessed in the solution and solid phases (Zümreoglu-Karan, 2006). Complexes with general formula $\text{M}^{n+}(\text{AscH}^-)_n \cdot x\text{H}_2\text{O}$ show good stability without any ET from Asch₂ to the metal center (Cesario *et al.*, 2017; Davies, 1992). The complex stability can affect the species distribution and, consequently, the availability for redox reactions.

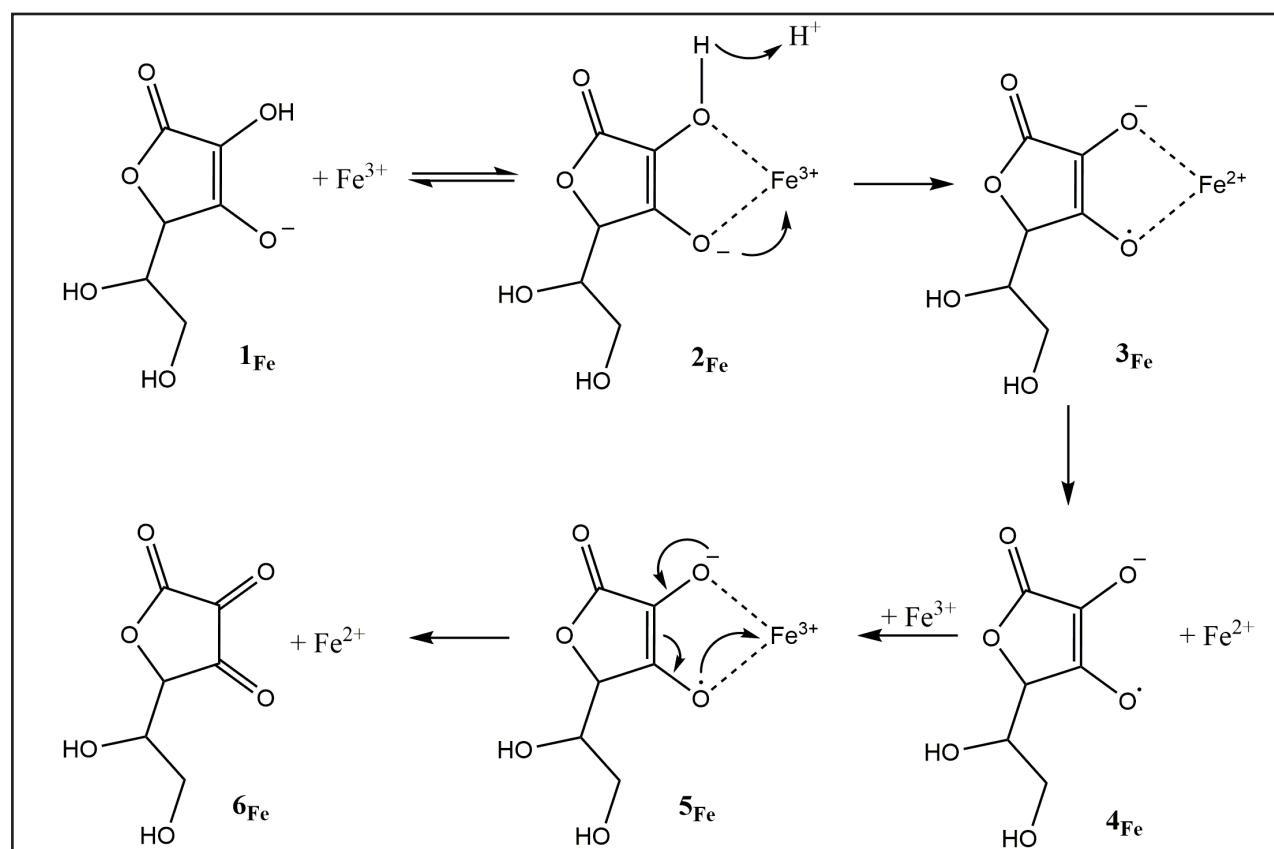
A recent study with cichoric acid evaluated the effects of metal complexation on the antioxidant activity of the molecule, via radical scavenging capacity. Results showed a slight increase in radical inhibition for complexes with redox inert metals (Na^+ and Zn^{2+}) (Swiderski *et al.*, 2020). Nonetheless, to the best of the authors' knowledge, the effect of complexation upon the antioxidant activity of Asch₂ has not yet been investigated.

3.3 Special cases, formation of catalytical systems: Cu(II) and Fe(III)

The E° for the redox pair $\text{Fe}^{3+} + \text{e}^- \rightleftharpoons \text{Fe}^{2+}$ is 0.77 V, which is relatively high and indicates that reduction of Fe^{3+} is thermodynamically favorable; however, it is unusual to find Fe^{2+} ions in oxidizing environment, as in the presence of O_2 . Fe^{3+} has a semi-

filled d orbital ($3d^5$), ensuring greater stability compared to Fe^{2+} ($3d^6$), especially when complexed with intermediate or weak field ligands which promote a smaller unfolding of the crystalline field, thus stabilizing the high spin species. The easiness with which Fe^{2+} is quickly oxidized to Fe^{3+} , in the presence of O_2 , provides a catalytic system that promotes $AsCH_2$ oxidation, even with trace amounts of Fe^{3+} in the medium (Kontoghiorghe *et al.*, 2020; Martell, 1982). Interaction of iron complexes with $AsCH_2$ form powerful catalysts for Fenton reactions which behave as pro-oxidants, as seen in the proposed mechanism illustrated in Figure 3 (Yuan *et al.*, 2021).

Figure 3 – Proposed mechanism for the catalytic oxidation of $AsCH_2$ by Fe^{3+}

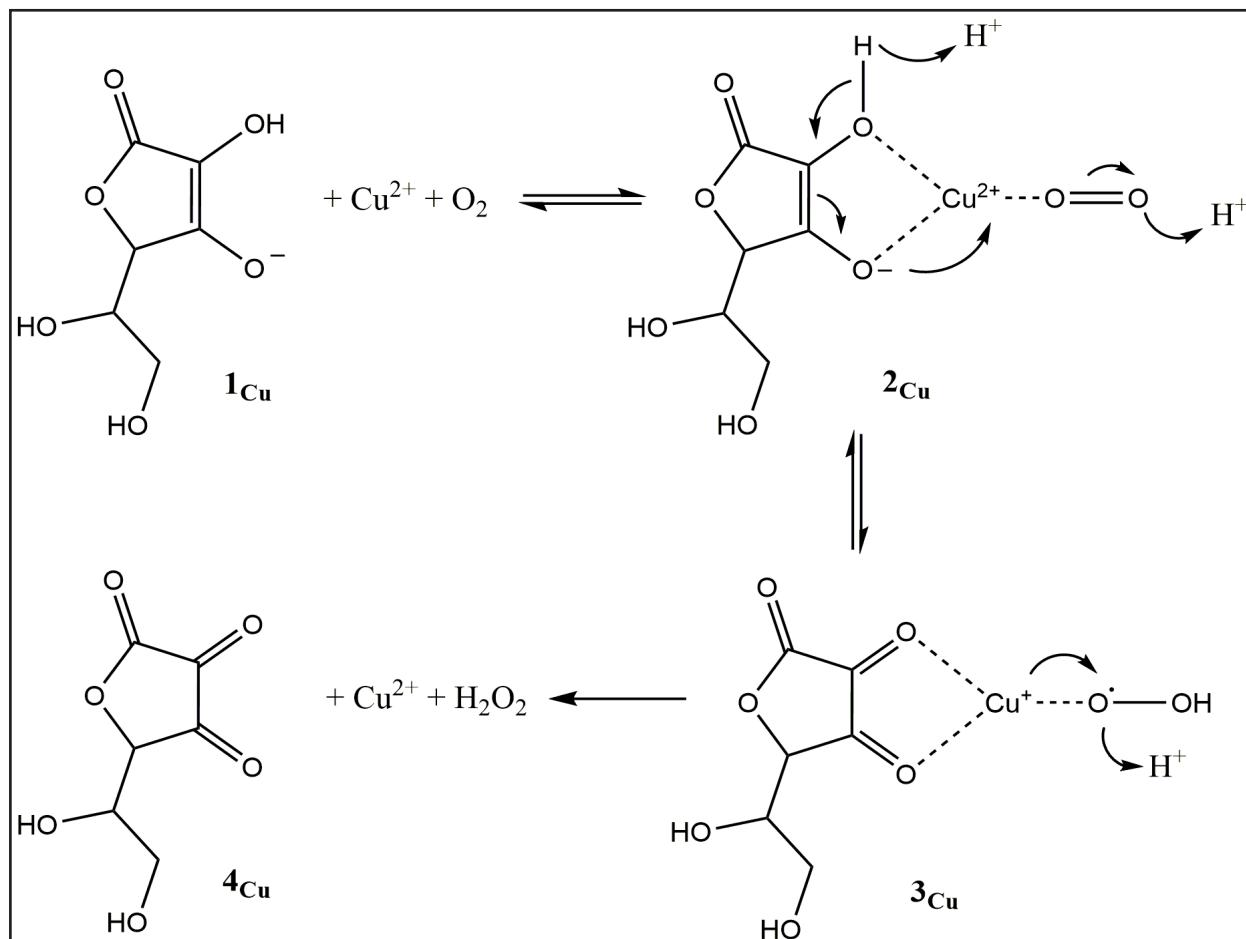


Source: Yuan *et al.*, 2021

The Cu^{2+} cation is a classic case of the Jahn-Teller effect with a greater stabilization of the d_{z^2} orbital in the $3d^9$ configuration. Cu^+ , in turn, has a full filled subshell ($3d^{10}$) which can also provide stability to the monovalent cation. Nevertheless, since it is a

soft acid (HSAB theory), Cu⁺ needs an equally soft base, little polarizable, as an S-donor base in order to form a stable complex. In an aqueous medium, the hydration enthalpy for Cu⁺ is so high that the cation is quickly oxidized to the divalent state (Khan and Martell, 1967). The redox behavior of Cu²⁺ in aqueous solution promotes the catalytic oxidation of AscH₂ with hydrogen peroxide formation (Martell, 1982), as seen in the proposed mechanism depicted in Figure 4.

Figure 4 – Proposed mechanism for the catalytic oxidation of AscH₂ by Cu²⁺, with hydrogen peroxide formation



Source: Martell, 1982

Table 1 – Selected standard reduction potentials for some transition metals (Lide, 2004)

Redox reaction	E°	Redox reaction	E°
$\text{Cu}^{2+} + e^- \rightleftharpoons \text{Cu}^+$	0.15	$\text{Co}^{3+} + e^- \rightleftharpoons \text{Co}^{2+}$	1.92
$\text{Fe}^{3+} + e^- \rightleftharpoons \text{Fe}^{2+}$	0.77	$\text{Co}^{2+} + e^- \rightleftharpoons \text{Co}$	-0.28
$\text{Ni}^{2+} + 2e^- \rightleftharpoons \text{Ni}$	-0.26	$\text{O}_2 + 2\text{H}^+ + 2e^- \rightleftharpoons \text{H}_2\text{O}_2$	0.69
$\text{Cr}^{3+} + e^- \rightleftharpoons \text{Cr}^{2+}$	-0.41	$\text{VO}_2^+ + 2\text{H}^+ + e^- \rightleftharpoons \text{VO}^{2+} + \text{H}_2\text{O}$	0.99
$\text{Al}^{3+} + 3e^- \rightleftharpoons \text{Al}$	-1.62	$\text{Cr}_2\text{O}_7^{2-} + 14\text{H}^+ + 6e^- \rightleftharpoons 2\text{Cr}^{3+} + 7\text{H}_2\text{O}$	1.36

4 CONCLUSION

Metal ions affect the reaction mechanism and antioxidant potentials. AsCH_2 , one of the most classic antioxidant agents studied in the literature, suffers a strong influence in the presence of metal ions forming stable metal complexes.

Based on this logic, it would be interesting to have more experimental tests in future works, mainly evaluating the complexation chemistry for AsCH_2 ; in spite of having been studied for a long time, there is no clear information in the literature with respect to characterization of crystals, for instance. In addition, there are no studies assessing the antioxidant activity of AsCH_2 complexes, especially regarding complexes with fixed oxidation states; those are known complexes, but their antiradical potentials have not yet been evaluated.

Interference of traces of metals is also very relevant, since the ability to act as a reducing agent can lead AsCH_2 to promote the reduction of Fe^{3+} to Fe^{2+} , for example, thus catalyzing Fenton reactions and resulting in an apparent pro-oxidant activity.

REFERENCES

ADACH, A.; JANYST, J.; CIEŚLAK-GOLONKA, M. Interaction of carcinogenic chromium(VI) Oxide, CrO_3 , with main nonenzymatic cellular reductants at physiological conditions. Electronic spectra and magnetic studies. **Spectroscopy Letters**, v. 28, n. 8, p. 1259–1273, 1995.

CESARIO, D.; FURIA, E.; MAZZONE, G.; BENEDUCI, A.; DE LUCA, G.; SICILIA, E. Complexation of Al³⁺ and Ni²⁺ by L-Ascorbic Acid: An Experimental and Theoretical Investigation. **J. Phys. Chem. A**, v. 121, n. 51, p. 9773–9781, 2017.

CRANS, D. C.; BARUAH, B.; GAIDAMAUSKAS, E.; LEMOS, B. Ç.; LORENZ, B. B.; JOHNSON, M. D. Impairment of ascorbic acid's anti-oxidant properties in confined media: inter and intramolecular reactions with air and vanadate at acidic pH. **Journal of Inorganic Biochemistry**, v. 102, n. 5–6, p. 1334–1347, 2008.

DAVIES, M. B. Reactions of L-ascorbic acid with transition metal complexes. **Polyhedron**, v. 11, n. 3, p. 285–321, 1992.

FERRER, E. G.; WILLIAMS, P. A. M.; BARAN, E. J. Interaction of the vanadyl(IV) cation with L-ascorbic acid and related systems. **Zeitschrift fur Naturforschung - Section B Journal of Chemical Sciences**, v. 53, n. 2, p. 256–262, 1998.

HVOSLEF, J. The Crystal Structure of L-Ascorbic Acid, "Vitamin C". **Acta Chemica Scandinavica**, v. 18, p. 841–842, 1964.

ISMAIL, A.; NAEEM, I.; GONG, Y. Y.; ROUTLEDGE, M. N.; AKHTAR, S.; RIAZ, M.; RAMALHO, L. N. Z.; OLIVEIRA, C. A. F. de; ISMAIL, Z. Early life exposure to dietary aflatoxins, health impact and control perspectives: A review. **Trends in Food Science & Technology**, v.112, p.212-224, 2021. DOI: <https://doi.org/10.1016/j.tifs.2021.04.002>

JABS, W.; GAUBE, W. Verbindungen der L-Ascorbinsäure mit Metallen. I. Zur Darstellung von Ascorbatkomplexen einiger 3d-Elemente. **ZAAC – Journal of Inorganic and General Chemistry**, v. 514, n. 7, p. 179–184, 1984.

JABS, W.; GAUBE, W. Verbindungen der L-Ascorbinsäure mit Metallen. IV. Ligandeigenschaften des Monoanions der L-Ascorbinsäure, C₆H₇O₆⁻. **ZAAC – Journal of Inorganic and General Chemistry**, v. 538, n. 7, p. 166–176, 1986.

JAGER, A. V.; TEDESCEO, M. P.; SOUTO, P. C. M. C.; OLIVEIRA, C. A. F. Assessment of aflatoxin intake in São Paulo, Brazil. **Food Control**, v.33, p.87-92, 2013. DOI: <https://doi.org/10.1016/j.foodcont.2013.02.016>

JAKŠIĆ, S.; BALOŠ, M. Ž.; POPOV, L.; KRSTOVIĆ, S. Optimisation, validation and comparison of methods for aflatoxin M1 determination in cheese. **International Journal of Dairy Technology**, v.74, p.681-688, 2021. DOI: <https://doi.org/10.1111/1471-0307.12784>

JASUTIENE, I.; GARMIENE, G.; KULIKAUŠKIENE, M. Pasteurisation and fermentation effects on aflatoxin M₁ stability. **Milchwissenschaft**, v.61, n.1, p.75-79. 2006. Available at: https://www.researchgate.net/publication/289151830_Pasteurisation_and_fermentation_effects_on_Aflatoxin_M1_stability. Accessed in: Sep. 21, 2023.

JECFA – Joint FAO/WHO Expert Committee on Food Additives, 2011. **Aflatoxin M1**. Available at: <https://inchem.org/documents/jecfa/jecmono/v47je02.htm>. Accessed in: Dec. 1st, 2021.

KHANEHGAH, A. M.; MOOSAVI, M.; OMAR, S. S.; OLIVEIRA, C. A. F.; KARIMI-DEHKORDI, M.; KAKHRI, Y.; HUSEYN, E.; NEMATOLLANI, A.; FARAHANI, M.; SANT'ANA, A. S. The prevalence and concentration of aflatoxin M1 among different types of cheeses: A global systematic review, meta-analysis, and meta-regression. **Food Control**, v.125, p.107960, 2021. DOI: <https://doi.org/10.1016/j.foodcont.2021.107960>

KHAN, M. M. T.; MARTELL, A. E. Metal ion and metal chelate catalyzed oxidation of ascorbic acid by molecular oxygen. II. Cupric and ferric chelate catalyzed oxidation. **Journal of the American Chemical Society**, v. 89, n. 26, p. 7104-7111, 1 dez. 1967.

KANNER, J.; MENDEL, H.; BUDOWSKI, P. Prooxidant and antioxidant effects of ascorbic acid and metal salts in a β-carotene-linoleate model system. **Journal of Food Science**, v. 42, n. 1, p. 60-64, 1 jan. 1977.

KAUR, S.; BEDI, J. S.; DHAKA, P.; VIJAY, D.; AULAKH, R. S. Exposure assessment and risk characterization of aflatoxin M1 through consumption of market milk and milk products in Ludhiana, Punjab. **Food Control**, v.126, p.107991, 2021. DOI: <https://doi.org/10.1016/j.foodcont.2021.107991>.

KONTOGHIORGHES, G. J.; KOLNAGOU, A.; KONTOGHIOTGHE, C. N.; MOUROUZIDIS, L. TIMOSHNIKOV, V. A.; POLYAKOV, N. E. Trying to Solve the Puzzle of the Interaction of Ascorbic Acid and Iron: Redox, Chelation and Therapeutic Implications. **Medicines**, v. 7, n. 8, p. 45, 2020.

KUIPER-GOODMAN, T. Uncertainties in the risk assessment of three mycotoxins: aflatoxin, ochratoxin, and zearalenone. **Canadian Journal of Physiology and Pharmacology**, v.68, p.1017-1024, 1990. DOI: <https://doi.org/10.1139/y90-155>

LIDE, D. R. CRC Handbook of Chemistry and Physics, 84th Edition. **Journal of the American Chemical Society**, v. 126, n. 5, p. 1586-1586, 2004.

LONDOÑO, V. A. G.; BOASSO, A. C.; PAULA, M. C. Z. de; GARCIA, L. P.; SCUSSEL, V. M.; RESNIK, S.; PACÍN, A. Aflatoxin M1 survey on randomly collected milk powder commercialized in Argentina and Brazil. **Food Control**, v.34, p.752-755, 2013. DOI: <https://doi.org/10.1016/j.foodcont.2013.06.030>

LOY, D. D.; LUNDY, E. L. Nutritional properties and feeding value of corn and its coproducts. In: **Corn: Chemistry and Technology**, AACC International Press, p. 633-659, 2019. DOI: <https://doi.org/10.1016/B978-0-12-811971-6.00023-1>

MARCONDES, M. M. **Incidência de podridão de colmo e grãos ardidos em híbridos de milho sob diferentes densidades de plantas e épocas de colheita**. Dissertação de mestrado. Universidade Estadual do Centro-Oeste. Guarapuava – PR. 2012. Available at: <https://1library.org/document/zw0vmlgy-prdissertacao-de-mestrado-marielle-martins.html>. Accessed in: Jan. 10, 2022.

MARTELL, A. E. Chelates of Ascorbic Acid. In: Ascorbic Acid: Chemistry, Metabolism, and Uses. **American Chemical Society**, 1982, v. 200, p. 153-178.

MARTINS, F. A.; FERREIRA, F. M. D.; FERREIRA, F. D.; BANDO, E.; NERILO, S. B.; HIROOKA, E. Y.; MACHINSKI JR., M. Daily intake estimates of fumonisins in corn-based food products in the population of Parana, Brazil. **Food Control**, v.26, n.2, p.614-618, 2012. DOI: <https://doi.org/10.1016/j.foodcont.2012.02.019>

MATOS, C. J.; SCHABO, D. C.; NASCIMENTO, Y. M. do; TAVARES, J. F.; LIMA E. de O.; CRUZ, P. O. da; SOUZA, E. L. de; MAGNANI, M.; MAGALHÃES, H. I. F. Aflatoxin M1 in Brazilian goat milk and health risk assessment. **Journal of Environmental Science and Health, Part B**, v.56, n.4, p.415-422, 2021. DOI: <https://doi.org/10.1080/03601234.2021.1892434>

MIN, L.; FINK-GREMMELS, J.; LI, D.; TONG, X.; TANG, J.; NAN, X.; YU, Z.; CHEN, W.; WANG, G. An overview of aflatoxin B1 biotransformation and aflatoxin M1 secretion in lactating dairy cows. **Animal Nutrition**, v.7, n.1, p.42-48, 2021. DOI: <https://doi.org/10.1016/j.aninu.2020.11.002>

MOLLAJUSEFIAN, I.; RANAEI, V.; PILEVAR, Z.; CABRAL-PINTO, M. M. S.; ROSTAMI, A.; NEMATOLAHI, A.; KHEDHER, K. M.; THAI, V. N.; FAKHRI, Y.; KHANEGHAH, A. M. The concentration of aflatoxin M1 in raw and pasteurized milk: A worldwide systematic review and meta-analysis. **Trends in Food Science & Technology**, v.115, p.22-30, 2021. DOI: <https://doi.org/10.1016/j.tifs.2021.06.033>

MOTTA, T. P.; FRIZZARIN, A.; MARTINS, T.; MIRANDA, M. S.; ARCARO, J. R. P.; AMBRÓSIO, L. A.; POZZI, C. R. Study on the occurrence of fungi and aflatoxina B1 in the diet of dairy cattle in São Paulo, Brazil. **Pesquisa Veterinária Brasileira**, v.35, p. 23-28, 2015. DOI: <https://doi.org/10.1590/S0100-736X2015000100006>

NACHTMANN, C.; GALLINA, S.; RASTELLI, M.; FERRO, G. L.; DECASTELLI, L. Regional monitoring plan regarding the presence of aflatoxin M1 in pasteurized and UHT milk in Italy. **Food Control**, v.18, n.6, p.623-629, 2007. DOI: <https://doi.org/10.1016/j.foodcont.2006.01.001>

NAVARRO, R. B.; ALMEIDA, R. de; POZZA, M. S. dos S.; BÁNKUTI, F. I.; ITAVO, C. B. C. F.; SCHENEEBERGER, C.; DIAS, A. M.; VITAL, A. C.; SANTOS, G. T. Dos. Presence of Mycotoxins in Feed and Dairy Products of Cattle in Paraná, Brazil. **Journal of Agricultural Studies**, 2020, v.8, p.505-514. DOI: <https://doi.org/10.5296/jas.v8i3.16856>

NGUYEN, T.; FLINT, S.; PALMER, J. Control of aflatoxin M1 in milk by novel methods: A review. **Food Chemistry**, v.311, p.125-984, 2020. DOI: <https://doi.org/10.1016/j.foodchem.2019.125984>

NGUYEN, T.; PALMER, J.; LOO, T.; SHILTON, A.; PETCU, M.; NEWSON, H. L.; FLINT, S. Investigation of UV light treatment (254 nm) on the reduction of aflatoxin M1 in skim milk and degradation products after treatment. **Food Chemistry**, v.390, p.133165, 2022. DOI: <https://doi.org/10.1016/j.foodchem.2022.133165>

NIMSE, S. B.; PAL, D. Free radicals, natural antioxidants, and their reaction mechanisms. **RSC Advances**, v. 5, n. 35, p. 27986–28006, 2015.

NYOKABI, S.; LUNING, P. A.; BOER, I. J. M. de; KORIR, L.; MUUNDA, E.; BEBE, B. O.; LINDAHL, J.; BETT, B.; OOSTING, S. J. Milk quality and hygiene: Knowledge, attitudes and practices of smallholder dairy farmers in central Kenya. **Food Control**, v.130, p.108303, 2021. DOI: <https://doi.org/10.1016/j.foodcont.2021.108303>

OLIVEIRA, C. A. F. de; SEBASTIÃO, L. S.; FAGUNDES, H.; ROSIM, R. E.; FERNANDES, A. M. Determinação de aflatoxina B1 em rações e aflatoxina M1 no leite de propriedades do Estado de São Paulo. **Food Science and Technology**, v.30, p.221-225, 2010. DOI: <https://doi.org/10.1590/S0101-20612010000500034>

OLIVEIRA, C. P., SOARES, N. de F. F.; OLIVEIRAM T. V. de; BAFFA JÚNIOR, J. C.; SILVA, W. A. da. Aflatoxin M1 occurrence in ultra high temperature (UHT) treated fluid milk from Minas Gerais/Brazil. **Food Control**, v.30, p.90-92, 2013. DOI: <https://doi.org/10.1016/j.foodcont.2012.07.026>

PEHLIVAN, F. E. Vitamin C: An Antioxidant Agent. In: **Vitamin C**. InTech, 2017.

PEREIRA, N.; FRANCESCHINI, S.; PRIORE, S. Qualidade dos alimentos segundo o sistema de produção e sua relação com a segurança alimentar e nutricional: revisão sistemática. **Saúde e Sociedade**, v.29, n.4, p.200031, 2020. DOI: <https://doi.org/10.1590/S0104-12902020200031>

PICININ, L. C. A.; CERQUEIRA, M. M. O. P.; VARGAS, E. A.; LANA, A. M. Q.; TOALDO, I. M.; BORDIGNON-LUIZ, M. T. Influence of climate conditions on aflatoxin M1 contamination in raw milk from Minas Gerais State, Brazil. **Food Control**, v.31, p.419-424, 2013. DOI: <https://doi.org/10.1016/j.foodcont.2012.10.024>

PIETRI, A.; FORTUNATI, P.; MULAZZI, A.; BERTUZZI, T. Enzyme-assisted extraction for the HPLC determination of aflatoxin M1 in cheese. **Food Chemistry**, v.192, p.235-241, 2016. DOI: <https://doi.org/10.1016/j.foodchem.2015.07.006>

PIRES, R. C.; PORTINARI, M. R. P.; MORAES, Z.; KHANEGBAH, A. M.; GONÇALVES, B. L.; ROSIM, R. E.; OLIVEIRA, C. A. F.; CORASSIN, C. H. Evaluation of Anti-Aflatoxin M1 effects of heat-killed cells of *Saccharomyces cerevisiae* in Brazilian commercial yogurts. **Quality Assurance and Safety of Crops & Foods**, v.14, n.1, p.75-81, 2022. DOI: <https://doi.org/10.15586/qas.v14i1.1006>

POUR, S. H.; MAKMOUDI, S.; MASOUMI, S.; REZAIE, S.; BARAC, A.; RANJBARAN, M.; OLIYA, S.; MEHRAVAR, F.; SASANI, E.; NOORBAKHSH, F.; KHODAVAISY, S. Aflatoxin M1 contamination level in Iranian milk and dairy products: A systematic review and meta-analysis. **World Mycotoxin Journal**, v.13, p.67-82, 2020. DOI: <https://doi.org/10.3920/WMJ2019.2485>

PRANDINI, A.; TANSINI, G. ; SIGOLO, S. ; FILIPPI, L. ; LAPORTA, M. ; PIVA, G. On the occurrence of aflatoxin M1 in milk and dairy products. **Food and Chemical Toxicology**, v.47, p.984-991, 2009. DOI: <https://doi.org/10.1016/j.fct.2007.10.005>

PRESTES, I. D.; ROCHA, L. O.; NUNES, K. V. M.; SILVA, N. C. C. Fungi and mycotoxins in corn grains and their consequences. **Scientia agropecuaria**, v.10, p.559-570, 2019. DOI: <https://doi.org/10.17268/sci.agropecu.2019.04.13>

PURCHASE, I. F. H.; STEYN, M.; RINISMA, R.; TUSTIN, R. C. Reduction of the aflatoxin M content of milk by processing. **Food and Cosmetics Toxicology**, v.10, n.383-387, 1972. DOI: [https://doi.org/10.1016/S0015-6264\(72\)80256-6](https://doi.org/10.1016/S0015-6264(72)80256-6)

QUEVEDO GARZA, P. A.; AMADOR-ESPEJO, G. G.; CANTÚ-MARTÍNEZ, P. C.; TRUJILLO-MESA, J. A. Aflatoxin M1 occurrence in fluid milk commercialized in Monterrey, Mexico. **Journal of Food Safety**, v.38, n.6, p.e12507, 2018. DOI: <https://doi.org/10.1111/jfs.12507>

RAMOS, C. E. C. de O.; DAMASCENO, J. C.; KAZAMA, R.; VIEIRA, T. S. W. J.; ZAMBOM, M. A.; FERREIRA, F. G.; SANTOS, G. T. dos. Seasonal milk contamination by aflatoxin M1, organophosphates and carbamates in Paraná-Brazil. Semina: **Ciências agrárias**, v.37, p.2145-2153, 2016. DOI: <https://doi.org/10.5433/1679-0359.2016v37n4p2145>

RITACCA, A. G. et al. Experimental and theoretical study of the complexation of Fe^{3+} and Cu^{2+} by L-ascorbic acid in aqueous solution. **Journal of Molecular Liquids**, v. 355, p. 118973, 2022.

ROUSSI, V.; GOVARIS, A.; VARAGOULI, A.; BOTSOGLOU, N. A. Occurrence of aflatoxin M₁ in raw and market milk commercializes in Greece. **Food Additives and Contaminants**, v.19, n.9, p.863-868, 2002. DOI: <https://doi.org/10.1080/02652030210146864>

SAFARI, N.; ARDAKANI, M. M.; HEMMATI, R.; PARRONI, A.; BECCACCIOLI, M.; REVERBERI, M. The Potential of Plant-Based Bioactive Compounds on Inhibition of Aflatoxin B1 Biosynthesis and Down-regulation of *aflR*, *aflM* and *aflP* Genes. **Antibiotics**, v.9, p.728, 2020. DOI: <https://doi.org/10.3390/antibiotics9110728>

ŞANLI, T.; DEVECI, O.; SEZGIN, E. Effects of pasteurization and storage on stability of aflatoxin M1 in yogurt. **Kafkas Üniversitesi Veteriner Facultesi Dergisi**, v.18, n.6, p.987-990, 2012. DOI: <https://doi.org/10.9775/KVFD.2012.6887>

SANTILI, A. B. N.; CAMARGO, A. C. de; NUNES, R. de S. R.; GLORIA, E. M. da; MACHADO, P. F.; CASSOLI, L. D.; DIAS, C. T. dos S.; CALORI-DOMINGUES, M. A. Aflatoxin M1 in raw milk from different regions of São Paulo state-Brazil. **Food Additives and Contaminants: Part B Surveillance**, v.8, p.207-214, 2015. DOI: <https://doi.org/10.1080/19393210.2015.1048538>

SANTOS, A. L.; BANDO, E.; JUNIOR, M. M. Ocorrência de aflatoxina M1 em leite bovino comercializado no estado do Paraná, Brasil. **Semina: Ciências agrárias**, v.35, p.371-374, 2014. DOI: <https://doi.org/10.5433/1679-0359.2014v35n1p371>

SANTOS, J. S.; GRANELLA, V.; PIGATTO, G. M.; REINIGER, L. R. S.; COSTABEBER, I. H. Aflatoxin M1 in pasteurized and raw milk from organic and conventional systems. **Journal für Verbraucherschutz und Lebensmittelsicherheit**, v.11, p.299-304, 2016. DOI: <https://doi.org/10.1007/s00003-016-1039-z>

SANTOS, J. S.; FRANÇA, V. R.; KATTO, S.; SANTANA, E. H. W. Aflatoxin M₁ in pasteurized, UHT milk and milk powder commercialized in Londrina, Brazil and estimation of exposure. **Archivos Latinoamericanos de Nutrición**, v.65, p.181-185, 2015. Available at: http://ve.scielo.org/scielo.php?script=sci_arttext&pid=S0004-06222015000300007. Accessed in: Dec. 1st, 2021.

SARTORI, A. V.; MATTOS, J. S. de; MORAES, M. H. P. de; NÓBREGA, A. W. da. Determination of aflatoxins M1, M2, B1, B2, G1, and G2 and ochratoxin A in UHT and powdered milk by modified QuEChERS method and ultra-high-performance liquid chromatography tandem mass spectrometry. **Food Analytical Methods**, v.8, p.2321-2330, 2015. DOI: <https://doi.org/10.1007/s12161-015-0128-4>

SIBAJA, K. V. M.; GARCIA, S. De O.; NOGUEIRA, W. V.; OLIVEIRA, F. K. de; BADIALE-FURLONG, E.; GARDA-BUFFON, J. Dietary exposure assessment of aflatoxin M1 in milk and dairy products of Latin America. **Food Reviews International**, v.38, p.669-682, 2022. DOI: <https://doi.org/10.1080/87559129.2021.1880434>

SILVA, I. M. de M.; CRUZ, A. G. da; ALI, S.; FREIRE, L. G. D.; FONSECA, L. M.; ROSIM, R. E.; CORASSIN, C. H.; OLIVEIRA, R. B. A. de; OLIVEIRA, C. A. F. de. Incidence and Levels of Aflatoxin M1 in Artisanal and Manufactured Cheese in Pernambuco State, Brazil. **Toxins**, v.15, n.3, p.182, 2023. DOI: <https://doi.org/10.3390/toxins15030182>

SILVA, M. V.; JANEIRO, V.; BANDO, E.; MACHINSKI JR., M. Occurrence and estimative of aflatoxin M₁ intake in UHT cow milk in Paraná State, Brazil. **Food Control**, v.53, p.222-225. 2015. DOI: <https://doi.org/10.1016/j.foodcont.2015.01.025>

SWIDERSKI, G.; JABLONSKA-TRYPUC, A.; KALINOWSKA, M.; SWISLOCKA, R.; KARPOWICZ, D.; MAGNUSZEWSKA, M.; LEWANDOWSKI, W. Spectroscopic, Theoretical and Antioxidant Study of 3d-Transition Metals (Co(II), Ni(II), Cu(II), Zn(II)) Complexes with Cichoric Acid. **Materials**, v. 13, n. 14, p. 3102, 2020.

TACO. **Tabela Brasileira de Composição de Alimentos**. Available at: https://www.cfn.org.br/wp-content/uploads/2017/03/taco_4_edicao_ampliada_e_revisada.pdf. Accessed in: Dec. 20, 2021.

TIMOSHNIKOV, V. A.; SELYUTINA, O. Y.; POLYAKOV, N. E.; DIDICHENKO, V.; KONTOGHIORGES, G. J. Mechanistic Insights of Chelator Complexes with Essential Transition Metals: Antioxidant/Pro-Oxidant Activity and Applications in Medicine. **International Journal of Molecular Sciences**, v. 23, n. 3, p. 1247, 2022.

TONON, K. M.; SAVI, G. D.; SCUSSSEL, V. M. Application of a LC-MS/MS method for multi-mycotoxin analysis in infant formula and milk-based products for young children commercialized in Southern Brazil. **Journal of Environmental Science and Health, Part B**, v.53, p.685-691, 2018. DOI: <https://doi.org/10.1080/03601234.2018.1474560>

TROMBETE, F. M.; CASTRO, I. M. de; TEIXEIRA, A. da S.; SALDANHA, T.; FRAGA, M. E. Aflatoxin M₁ contamination in grated parmesan cheese marketed in Rio de Janeiro-Brazil. **Brazilian Archives of Biology and Technology**, v.57, p. 269-273, 2014. DOI: <https://doi.org/10.1590/S1516-89132013005000015>

TU, Y. J.; NJUS, D.; SCHLEGEL, H. B. A theoretical study of ascorbic acid oxidation and HOO[·]/O₂^{·-} radical scavenging. **Organic and Biomolecular Chemistry**, v. 15, n. 20, p. 4417–4431, 2017.

USDA – Department of Agriculture and Department of Health and Human Services. **Dietary Guidelines for Americans**. Available at: <https://health.gov/our-work/nutrition-physical-activity/dietary-guidelines/previous-dietary-guidelines/2010>. Accessed in: Dec. 20, 2021.

VARIANE, A. C. F.; SANTOS, F. C. dos; CASTRO, F. F. De; BARBOSA-TESSMANN, I. P.; SANTOS, G. T. dos; POZZA, M. S. dos S. The occurrence of aflatoxigenic Aspergillus spp. in dairy cattle feed in Southern Brazil. **Brazilian Journal of Microbiology**, v.49, p.919-928, 2018. DOI: <https://doi.org/10.1016/j.bjm.2018.05.005>

VENÂNCIO, R. L.; LUDOVICO, A.; SANTANA, E. H. W. de; TOLEDO, E. A. de; REGO, F. C. de A.; SANTOS, J. S. dos. Occurrence and seasonality of aflatoxin M1 in milk in two different climate zones. **Journal of the Science of Food and Agriculture**, v.99, p.3203-3206, 2018. DOI: <https://doi.org/10.1002/jsfa.9487>

VILELA, D.; RESENDE, J. C. de; LEITE, J. B.; ALVES, E. The evolution of milk in Brazil in five decades. **Revista de Política Agrícola**, v.26, n.1, p.20, 2017. Available at: <https://seer.sede.embrapa.br/index.php/RPA/article/view/1243/1037>. Accessed in: Dec. 1st, 2021.

WHO – World Health Organization. **IARC Monographs on the Identification of Carcinogenic Hazards to Humans**, 2022. Available at: <https://monographs.iarc.who.int/list-of-classifications>. Accessed in: Aug. 29, 2022

YIANNIKOURIS, A.; JOUANY, J. P. Mycotoxins in feeds and their fate in animals: a review. **Anim. Res.**, v.51, p.81-89, 2002. DOI: <https://doi.org/10.1051/animres:2002012>

YIN, X.; CHEN, K.; CHENG, H.; CHEN, X.; FENG, S.; SONG, Y.; LIANG, L. Chemical Stability of Ascorbic Acid Integrated into Commercial Products: A Review on Bioactivity and Delivery Technology. **Antioxidants**, v. 11, n. 1, p. 153, 2022.

YUAN, D.; ZHANG, C.; TANG, S.; WANG, Z.; SUN, Q.; ZHANG, X.; JIAO, T.; ZHANG, Q. Ferric ion-ascorbic acid complex catalyzed calcium peroxide for organic wastewater treatment: Optimized by response surface method. **Chinese Chemical Letters**, v. 32, n. 11, p. 3387–3392, 2021. ZÜMREOGLU-KARAN, B. The coordination chemistry of Vitamin C: An overview. **Coordination Chemistry Reviews**, v. 250, n. 17, p. 2295–2307, 2006.

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