



Chemistry

Nanoemulsions of plant-based bioactive compounds with antimicrobial applications: a review

Nanoemulsões de compostos bioativos de base de plantas com aplicações antimicrobianas: uma revisão

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ABSTRACT

The search for alternative antimicrobial agents is attracting increasing scientific interest. Natural products of plant origin are sources of several substances with proven biological activities, including antimicrobial activity. The encapsulation of these products in the form of a nanoemulsion seeks to overcome problems inherent to these products, such as instability and degradation. Based on these considerations, we carried out a bibliographical survey of nanoemulsions produced from plant-derived substances, such as essential oils and extracts, with antimicrobial potential, focusing on antibacterial, antifungal and antiviral activities. Articles and documents published in scientifically relevant journals, as well as keywords classified from Health Sciences Descriptors, were used. All documents relevant to this search reported that nanoemulsions loaded with essential oils and plant extracts from different botanical species had in vitro antimicrobial activity against different microorganisms of medical importance, in addition to enhancing the antimicrobial effects of these bioproducts. Therefore, nanostructured antimicrobials with essential oils and plant extracts can be considered treatment options for microbial diseases: due to their physicochemical properties, they act as better delivery vehicles for natural products with good bioavailability, by reducing toxicity and prolonging the useful life of these natural antimicrobials, thus enhancing treatment for infectious human diseases.

Keywords: Antimicrobial activity; Nanoparticles; Alternative products

RESUMO

A busca por agentes antimicrobianos alternativos vem atraindo crescente interesse científico. Os produtos naturais de origem vegetal são fontes de diversas substâncias com atividades biológicas comprovadas, incluindo atividade antimicrobiana. O encapsulamento desses produtos na forma de nanoemulsão busca superar problemas inerentes a esses produtos, como instabilidade e degradação. Com base nessas considerações, realizamos um levantamento bibliográfico de nanoemulsões produzidas a partir de substâncias derivadas de plantas, como óleos essenciais e extratos, com potencial antimicrobiano, com foco nas atividades antibacteriana, antifúngica e antiviral. Foram utilizados artigos e documentos publicados em periódicos de relevância científica, bem como palavras-chave classificadas nos Descritores em Ciências da Saúde. Todos os documentos relevantes para esta pesquisa relataram que as nanoemulsões carregadas com óleos essenciais e extratos vegetais de diferentes espécies botânicas apresentaram atividade antimicrobiana *in vitro* contra diferentes microrganismos de importância médica, além de potencializar os efeitos antimicrobianos desses bioprodutos. Portanto, antimicrobianos nanoestruturados com óleos essenciais e extratos de plantas podem ser considerados opções de tratamento para doenças microbianas: devido às suas propriedades físico-químicas, atuam como melhores veículos de entrega de produtos naturais com boa biodisponibilidade, reduzindo a toxicidade e prolongando a vida útil desses antimicrobianos naturais, melhorando assim o tratamento de doenças humanas infecciosas.

Palavras-chave: Atividade antimicrobiana; Nanopartículas; Produtos alternativos

1 INTRODUCTION

Infectious diseases occupy fourth place in the ranking of diseases that caused the most hospitalizations and deaths in the world between 2000 and 2019 (Paho 2020). This fact is directly related to the presence of infectious microorganisms (bacteria, fungi, protozoa and viruses) that cause damage to the host (Madigan *et al.* 2016; Tortora *et al.* 2016). These diseases are generally treated with antimicrobials, substances of natural origin (antibiotics) or synthetic (chemotherapeutics), which act on the microorganisms, inhibiting their growth or causing their death (Sáez-Llorens *et al.* 2000).

Despite the need to use antibiotic therapy to treat infections, there are risks to the health of patients related to the high toxicity of these drugs, some of which are nephrotoxic (Kaloyanides, 1994), hepatotoxic or ototoxic (Oun *et al.* 2018). They also have a narrow therapeutic window and high teratogenic potential (Kaleelullah;

Garugula, 2021). In addition to these drugs' toxicity, another rising problem is microbial resistance, in which these microorganisms develop mechanisms that inactivate the active principle responsible for the antimicrobial action, which may be related to the action of enzymes, changes in the binding sites, physical barriers or action of efflux pumps (Reygaert, 2018; Kember *et al.* 2022).

Due to these drawbacks of antimicrobial therapy, the need has arising to develop new bioactive substances by molecular alteration of existing drugs or the discovery of new substances with antimicrobial potential (Spellberg *et al.* 2015). Among these new substances, the standouts are crude plant-derived substances (isolated by extractive methods) or substances that actively participate in green synthesis processes (Galie *et al.* 2018).

The effects of active ingredients of plant origin can vary according to the extraction method used, with emphasis on essential oils, aqueous extracts and ethanolic extracts. These methods are responsible obtaining various chemical compounds from the secondary metabolism of plants (Wallace, 2004). Secondary metabolites have various therapeutic activities, such as antimicrobial and antioxidant (Yang *et al.* 2018; Isah, 2019).

The encapsulation of these plant compounds in the form of an emulsion has strategic advantages by improving kinetic stability, preserving organoleptic characteristics, and enhancing penetration power, thus enhancing the desired effect (Chinnaiyan *et al.* 2022; Sahu *et al.* 2018). In addition, they promote greater bioavailability, increase solubility in water of poorly soluble substances and allow their controlled release (Karim *et al.* 2022).

Therefore, we carried out a bibliographic survey of nanoemulsions produced from active ingredients of plant origin, such as essential oils and extracts, with antimicrobial potential, focusing on antibacterial, antifungal and antiviral activities. The results can contribute to new studies to identify efficient nanoemulsions that can be applied to treat infectious diseases. The information reported here fills in the gaps in knowledge and makes these substances more readily available for future applications in the field of health, thus overcoming the limitations of conventional antimicrobials.

The bibliographic survey was carried out between June and November 2022, in the databases Google Scholar (<<https://scholar.google.com.br>>), PUBMED (<<https://pubmed.ncbi.nlm.nih.gov/>>), SciELO (<<https://www.scielo.org/>>), Science Direct (<<https://www.sciencedirect.com/>>), Scopus® (<<https://www.scopus.com/>>), LILACS (<<https://lilacs.bvsalud.org/>>) and BDTD (<<https://bdt.d.ibict.br/>>). To search for articles, keywords were used that were classified from the Health Sciences Descriptors (<<https://decs.bvsalud.org/>>): “Nanoemulsion”, “Polymeric Nanoparticles”, “Lipid Nanoparticles”, “Biological Products”, “Anti-Infective Agents”, “Anti-Bacterial Agents”, “Antifungal Agents” and “Antiviral Agents”, for sample refinement. Articles that addressed the topic in question, in Portuguese or English, published in journals present in the platforms mentioned above between 2012 to 2022 were selected for reading in full.

2 DEVELOPMENT

2.1 Natural products of plant origin

Extracts, isolates and oils extracted from natural sources are widely used for biomedical purposes, since many of them have antimicrobial properties (Boire *et al.* 2013; Samber *et al.* 2015; Mostafa *et al.* 2018; D’agostino *et al.* 2019). Although natural products have generally presented good results for application in health care, their use faces some limiting factors, such as sensitivity to physical changes, since they are vulnerable to heat, humidity and oxygen. Among the resulting problems are hydrolysis and oxidation reactions, causing structural changes (Bhargava *et al.* 2015; Ghani *et al.* 2018; Bedoya-Serna *et al.* 2018).

Therefore, they need greater care in their storage, packaging and transport, to assure there are no changes in their properties. In order to protect bioactive compounds, nanotechnology can be applied, with benefits such as bioavailability, protection against degradation, increased apparent solubility, and better physical and chemical biocompatibility of the active principle, thus enhancing the pharmacological action and protecting against toxicity (Rao; Khanum, 2016; Bazana *et al.* 2019).

Nanoencapsulated bioactive compounds offer various advantages, such as increased physicochemical stability, better antimicrobial activity, and minimization of side effects (Herculano *et al.* 2015; Donsi; Ferrari, 2016).

Encapsulation is a technique widely used in the pharmaceutical, chemical, food and biomedical sectors, in order to promote protection and make products of natural origin more manageable in formulations. Among the encapsulation techniques, the formation of nanoemulsions stands out for the production of drugs, by providing significant protection of these bioactive compounds (Bhargava *et al.* 2015; Ghani *et al.* 2018).

2.2 Nanoemulsions

Nanoemulsions are dispersions of two immiscible liquids stabilized by a surfactant, where the size of the dispersed droplets is on a nanometric scale (20 to 500 nm). A nanoemulsion's transparency is directly related to the diameter of the dispersed droplets (translucent < » 200 nm > milky) (Quintão *et al.* 2013; Nascimento *et al.* 2020). Figure 1 represents the behavior of three nanoemulsions with different diameters of dispersed oily particles.

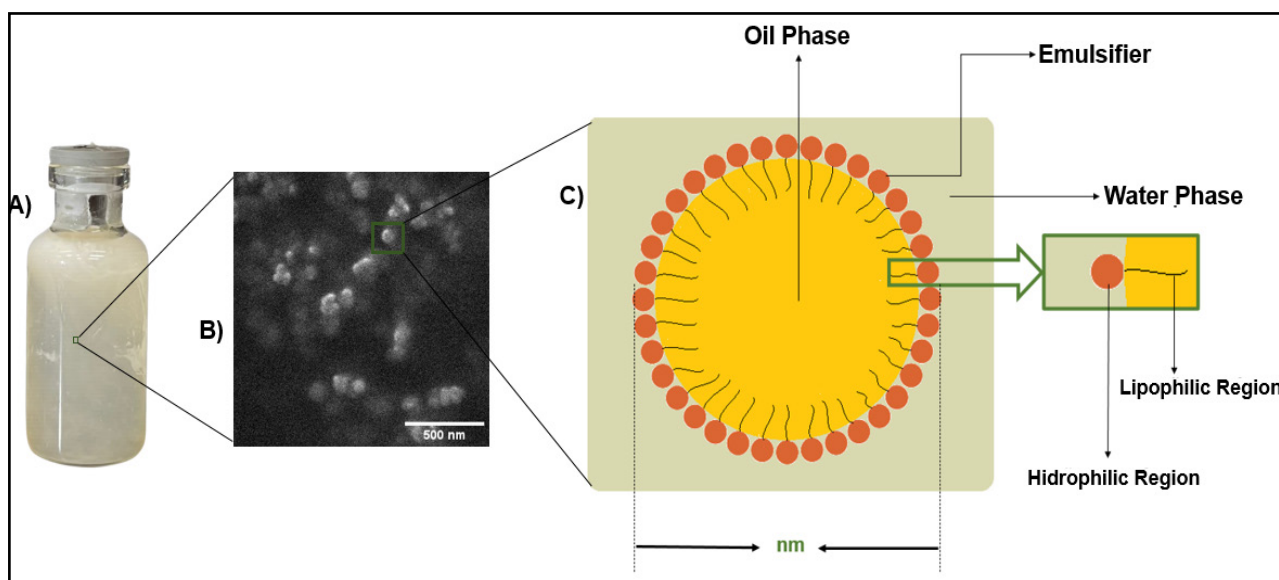
Figure 1 - Nanoemulsions and their behavior against the different diameters of their particles



Source: Authors' private collection (February 2023)

Nanoemulsions are classified as water-in-oil (W/O) or oil-in-water (O/W), depending on the hydrophilicity or lipophilicity of the dispersing medium (Wang *et al.* 2007). The surfactant acts as a stability barrier of nanoemulsions, characterized as an amphiphilic compound that is positioned between the two phases of the emulsion (W/O or O/W), creating an interfacial film that stabilizes the system (Kumar; Mandal, 2018). Figure 2 represents the layout of an O/W nanoemulsion system with the structure of each component, where it is possible to observe the aqueous phase, oil phase and emulsifying agent.

Figure 2 – Representative schematic of oil-in-water nanoemulsion. A) Macroscopic view of nanoemulsion (O/W); B) Nanoemulsion scanning electron microscopy; C) Schematic representation of a micelle, containing the oil phase inside, surrounded by the surfactant



Source: Authors' private collection (February 2023).

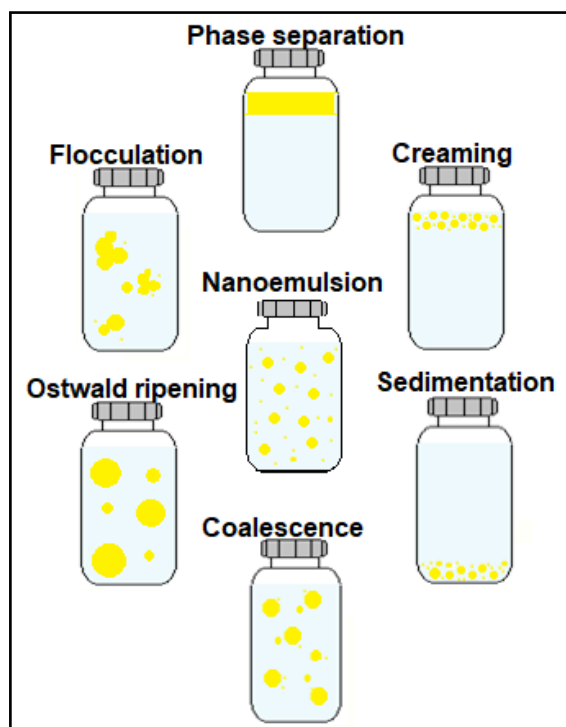
The addition of texture modifiers such as polymeric matrices of natural origin to the aqueous phase has often been used to improve the stability of nanoemulsions, since they have surfactant properties, thus possessing emulsifying and thickening capacity. They also coat the individual droplets, reducing their movement and avoiding the destabilization of emulsions and nanoemulsions due to the decrease of

the gravitational movement of the oil droplets. This delays or prevents the droplets' coalescence due to the increased viscosity of the aqueous phase (Guerra-Rosas *et al.* 2016; Maphosa; Jideani, 2018).

Emulsifiers or surfactants are fundamental to prevent the aggregation of nanoparticles, even after prolonged storage, by promoting the physicochemical stability of the nanoemulsion (Surassmo *et al.* 2010). Surfactants are amphiphilic substances and have a polar (hydrophilic) and a nonpolar (lipophilic) region, an ideal characteristic for the formation of O/W or W/O nanoparticles (Botelho *et al.* 2018).

Nanoemulsions have good stability of suspended nanoparticles, and the appearance of these systems is strongly related to the size of the formed particles (Weiss *et al.* 2009). The physical destabilization of emulsions is related to the spontaneous tendency to occupy the minimum interfacial space between their immiscible phases (Anton *et al.* 2008). Figure 3 shows the possible types of physicochemical instabilities related to nanoemulsion systems.

Figure 3 – Representative schematic of nanoemulsion instability



Source: Authors' private collection (February 2023)

Among the destabilization processes, creaming stands out. It is more likely to occur in O/W nanoemulsions, while sedimentation is more prevalent in W/O nanoemulsions (Zhang; McClements, 2018). Creaming is the formation of a creamy layer on the surface of the emulsion, which occurs due to the difference in density of the nanoemulsion components and the presence of forces external to the system, such as gravitational and centrifugal forces (Reddy *et al.* 1981; Rabinovich *et al.* 2004). In both sedimentation and creaming, the difference in the density of the materials and their low solubility in an aqueous medium occur (Tadros, 2006).

Coalescence, flocculation and Ostwald ripening occur due to partial solubilization of the oil phase in the aqueous phase, thus forming larger oil droplets dispersed in the system, which can lead to total phase separation. Flocculation can also be understood as the aggregation between the oil droplets, which consequently can lead to the formation of larger droplets, characterized as coalescence. Ostwald ripening, on the other hand, consists of an increase in the droplet size and a decrease in the total number of dispersed droplets in the system (Tadros, 1996; Franzol, Rezende, 2015).

2.3 Antimicrobial potential of nanoemulsions produced with plant-based bioactive compounds

The nanoemulsions synthesized with different essential oils and plant extracts have been tested *in vitro* against different microorganisms of medical importance, such as bacteria, viruses and fungi, as shown in Table 1. It summarizes the main reported findings regarding the active plant components, dispersing medium, surfactant, particle size of the nanoemulsions and the species of microorganisms.

Table 1 – Nanoemulsions produced from different plant extracts with antibacterial, antifungal and antiviral properties, in addition to presenting the dispersant medium, surfactant, particle size and the potentiated effect of nanoemulsions

(Continue)

Plant Antimicrobial	Dispersant/ Surfactant	Size/ Potentiation	Antimicrobial Activity	Ref.
<i>Allium sativum</i> Oil	Water/ Tween 80	36,6 nm / -	<i>Staphylococcus aureus</i> , <i>Escherichia coli</i> .	Hassan; Mujtaba (2019)
<i>Artemisia annua</i> Oil	Water/ Tweem 80	130-160 nm / -	<i>E. coli</i> , <i>S. aureus</i> , <i>Bacillus subtilis</i> , <i>Pseudomonas aeruginosa</i> , <i>Streptococcus pyogenes</i> , <i>Schizosaccharomyces pombe</i> , <i>Candida albicans</i> , <i>C. tropicalis</i> , <i>C. dubliniensis</i> , <i>C. krusei</i> .	Das et al. (2020)
<i>Azadirachta indica</i> Oil	Water/ Tween 20	70 nm/ +	<i>Vibrio vulnificus</i>	Jerobin et al. (2015)
<i>Cinnamomum spp.</i> Oil	Water/ Tweem 80	2040-40,4 nm/ -	<i>E. coli</i> , <i>S. aureus</i>	Valizadeh et al. (2018)
<i>Citrus limonum</i> Oil	Water/ Tweem 80	181,5 nm/ +	<i>S. aureus</i> , <i>Klebsiella pneumoniae</i> , <i>Salmonella typhimurium</i> , <i>Enterococcus faecalis</i> , <i>Photobacterium damsela</i> , <i>Vibrio vulnificus</i> , <i>Proteus mirabilis</i> , <i>Serratia liquefaciens</i> , <i>Pseudomonas luteola</i>	Yazgan et al. (2019)
<i>Citrus paradisi</i> Oil	Water/ Tweem 80	173,9-0,105 nm/ +	<i>E. coli</i> , <i>S. aureus</i>	Wang et al. (2022)
<i>Copaifera langsdorffii</i> Oil	Water/ Tweem 80	641,3-29,98 nm/ -	<i>Paracoccidioides lutzii</i> , <i>P. brasiliensis</i> , <i>P. americana</i> , <i>P. restrepiensis</i>	Silva et al. (2020)
<i>Croton cajucara</i> Oil	Water/ Pluronic	45,56 nm/ -	<i>Absidia cylindrospora</i> , <i>Cunninghamella elegans</i> , <i>Mucor circinelloide</i> , <i>M. muceno</i> , <i>M. ramosissimo</i> , <i>Rhizopus microsporu</i> , <i>R. oryzae</i> , <i>Syncephalastrum racemosum</i> , <i>C. albicans</i>	Azevedo et al. (2021)
<i>Cymbopogon martini</i> Oil	Water/ Tween 80	118-12 nm/ +	<i>E. faecalis</i>	Marinkovic et al. (2022)

^(*)Enhanced antimicrobial effect of plant products in the form of nanoemulsions; ^(*)Effects not potentiated or indifferent in the form of nanoemulsion.

Source: Authors' (February 2023)

Table 1 – Nanoemulsions produced from different plant extracts with antibacterial, antifungal and antiviral properties, in addition to presenting the dispersant medium, surfactant, particle size and the potentiated effect of nanoemulsions

(Continue)

Plant Antimicrobial	Dispersant/ Surfactant	Size/ Potentiation	Antimicrobial Activity	Ref.
<i>Eucalipto globulus Oil</i>	Sorbitan monooleate/ Tween 80	100 nm/ -	<i>P. aeruginosa, C. albicans, C. tropicalis, C. glabrata</i>	Quatrin <i>et al.</i> (2017)
<i>Jasminum humile L. Oil</i> <i>Jasminum grandiflorum L. Oil</i>	Water/ Tween 80	12-8 nm/ -	HAV (Virus Hepatite A), HSV (Herpes Virus)	Mansour <i>et al.</i> (2022)
<i>Lippia sidoides Cham Oil</i>	Compritol 888/ Sodium Odecyl Sulfate	445,5-213,1/ +	<i>Candida auris</i>	Baldirim <i>et al.</i> (2022)
<i>Mauritia flexuosa Lf. Total Oil</i>	Water/ Tween 80	270-196 nm/ -	<i>E. coli</i>	Leão <i>et al.</i> (2019)
<i>Melaleuca alternifolia Oil</i>	Water/ Tween 80	500-100 nm/ -	<i>E. coli</i>	Wei <i>et al.</i> (2022)
<i>Mentha pulegium Oil</i>	Water/ Tween 80	125 nm/ -	<i>E. coli, S. typhi, S. aureus, Listeria monocytogenes</i>	Damani <i>et al.</i> (2022)
<i>Mentha spicata L. Oil</i>	Water/ Tween 80	51,46 nm/ +	<i>S. typhimurium, S. aureus, E. coli, Bacillus cereus, L. monocytogenes</i>	Zamaniahari <i>et al.</i> (2022)
<i>Poiretia latifolia Oil</i>	Water/ Tween 80	-	<i>C. albicans, Candida krusei, C. tropicalis, C. glabrata, M. canis, Microsporium gypseum, T. rubrum, T. mentagrophyte</i>	Paim <i>et al.</i> (2018)
<i>Satureja hortensis Oil</i>	Water/ Tween 80	219 nm/ -	<i>K. pneumoniae, P. aeruginosa, Serratia marcescens, E. coli, S. typhi, S. aureus, L. monocytogenes, Staphylococcus haemolyticus</i>	Maccelli <i>et al.</i> (2020)
<i>Syzygium aromaticum Oil</i>	Water/ Tween 80	-	<i>A. niger, Colletotrichum gloeosporioides, Penicillium chrysogenum.</i>	Lima <i>et al.</i> (2021)
<i>Stenachaenium megapotamicum Oil</i>	Water/ Tween 80	210 nm/ +	<i>Epidermophyton floccosum, Scytalidium dimidiatum, T. rubrum</i>	Danielli <i>et al.</i> (2013)

⁽⁺⁾Enhanced antimicrobial effect of plant products in the form of nanoemulsions; ⁽⁻⁾Effects not potentiated or indifferent in the form of nanoemulsion.

Source: Authors' (February 2023)

Table 1 – Nanoemulsions produced from different plant extracts with antibacterial, antifungal and antiviral properties, in addition to presenting the dispersant medium, surfactant, particle size and the potentiated effect of nanoemulsions

(Continue)

Plant Antimicrobial	Dispersant/ Surfactant	Size/ Potentiation	Antimicrobial Activity	Ref.
<i>Thymus vulgaris</i> Oil	Water/ Tween 80	127 nm/ +	<i>C. albicans</i> , <i>C. glabrata</i> , <i>Candida parapsilosis</i> , <i>A. fumigatus</i> , <i>T. tonsurans</i> , <i>T. rubrum</i> , <i>T. mentagrophytes</i> , <i>M. canis</i> <i>Staphylococcus</i> sp., <i>Enterococcus</i> sp., <i>Escherichia</i> sp., <i>Pseudomonas</i> sp.	Moazeni <i>et al.</i> (2021) Sienkiewicz <i>et al.</i> (2012)
<i>Myrtus commnis</i> L. Oil	Water/ Tween 80	179 nm/ +	<i>E. coli</i> , <i>S. aureus</i>	Roozitalab <i>et al.</i> (2022)
<i>Matricaria chamomilla</i> L. Oil	Water/ Tween 80	20 nm/ +	<i>E. coli</i> , <i>P. aeruginosa</i> , <i>B. subtilis</i> , <i>S. aureus</i> , <i>S. pyogenes</i> , <i>S. pombe</i> , <i>C. albicans</i> , <i>C. tropicalis</i>	Das <i>et al.</i> (2019)
<i>Capsicum annum</i> L. Ethanol Extract	Water/ Tweem 80	-	<i>E. coli</i> , <i>S. aureus</i> , <i>C. albicans</i> , <i>Aspergillus niger</i> .	El-Naggar <i>et al.</i> (2020)
<i>Cuphea ígnea</i> Ethanol Extract	Water/ Tween 80	119 nm/ +	SARS CoV-2 (COVID-19)	Mahmoud <i>et al.</i> (2021)
<i>Curcuma spp.</i> Extract	Water/ Cremophor RH40	40 nm/ +	DENG-1, DENG-2, DENG-3, DENG-4 (DENGUE VÍRUS)	Nabila <i>et al.</i> (2020)
<i>Pimpinella anisum</i> Oil	Water/ Tween 80	117-275 nm/ +	<i>E. coli</i> , <i>L. monocytogenes</i>	Topuz <i>et al.</i> (2016)
<i>Pimpinella anisum</i> Ethanol Extract	Water/ Tween 80	400 nm/ +	<i>S. aureus</i> , <i>B. cereus</i> , <i>L. monocytogenes</i> , <i>E. coli</i> , <i>S. typhimurium</i> , <i>P. aeruginosa</i> , <i>Yersinia enterocolitica</i>	Ghazy <i>et al.</i> (2021)
<i>Ginkgo biloba</i> Ethanol Extract	Water/ Tween 80	97 nm/ +	H3N2 (Vírus influenza), HBV (Hepatitis B vírus)	Wang <i>et al.</i> (2015)

(^a)Enhanced antimicrobial effect of plant products in the form of nanoemulsions; (^b)Effects not potentiated or indifferent in the form of nanoemulsion.

Source: Authors' (February 2023)

Table 1 – Nanoemulsions produced from different plant extracts with antibacterial, antifungal and antiviral properties, in addition to presenting the dispersant medium, surfactant, particle size and the potentiated effect of nanoemulsions

				(Conclusion)
Plant Antimicrobial	Dispersant/ Surfactant	Size/ Potentiation	Antimicrobial Activity	Ref.
<i>Coriandrum sativum</i> Oil	Chitosan/ Tween 80	80-57 nm/ +	<i>A. Niger, Aspergillus fumigatus, A. sydowii, A. repens, A. versicolor, A. luchuensis, Alternaria alternata, Penicillium italicum, P. crysogenum, P. spnulosum, Mycelia sterilia, Clasoaporium herbarum, Fusarium poae, F. oxysporum.</i>	Das et al. (2019)
<i>Eucalipto citriodora</i> Oil	Chitosan/ Tween 80	1271-388 nm/ +	<i>S. aureus, S. typhimurium</i>	Abreu et al. (2020)
<i>Ocimum gratissimum</i> Linn. Oil	Cashew gum/ Tween 80	500-10 nm/ +	<i>E. coli, S. aureus, Salmonella enterica, Mycobacterium smegmatis, Mycobacterium bovis,</i>	Okonkwo et al. (2020)
<i>Psidium guajava</i> L.Oil	Chitosan/ Tween 80	96-40 nm/ +	<i>K. pneumoniae</i>	Zhang et al. (2020)
<i>Phyllanthus niruri</i> Methanolic Extract	Sodium Alginate/ Tween 80	192 nm/ +	<i>E. coli, K. pneumonia, S. aureus. S. typhimurium, Streptococcus mutans, B. subtilis, Trichophyton rubrum, Trichophyton mentagrophytes.</i>	Pathania et al. (2022)

(⁺)Enhanced antimicrobial effect of plant products in the form of nanoemulsions; (⁰)Effects not potentiated or indifferent in the form of nanoemulsion.

Source: Authors' (February 2023)

Among the studies analyzed in this review, 91.34% involved nanoemulsions produced with water as the aqueous phase, while the remaining 8.66% involved natural polymer matrices, with chitosan being most prevalent in the formulations. Among the hydrophilic surfactants, Tween 80 is the predominant one, due to its excellent solubility of essential oils and water miscibility. Among the most frequent natural products, essential oils stood out (91.34%), although ethanolic and hydroalcoholic extracts (8.66%) were also observed as active principles in some formulations.

Essential oils are aromatic compounds of natural origin with recognized biological activities. They are used as flavoring additives in foods and stabilizers in cosmetics. Their use as medicinal agents is variously due to their insecticidal, antioxidant, anti-inflammatory, anti-allergic and anticancer activities. Many essential oils also have antibacterial, antiviral and antifungal activities, stimulating their application also as natural antimicrobials in foods and beverages (Donsì; Ferrari, 2016).

Over the years, a plethora of research has shown the potent *in vitro* antimicrobial activity of various essential oils and their constituents against a wide range of microorganisms. However, as discussed earlier, the use of essential oils in biomedical systems is challenging due to their low stability and sensitivity to physical changes, since they are highly vulnerable to heat, moisture and oxygen. These problems can be overcome by encapsulating essential oils in suitable delivery systems, helping to improve their stability and biological activity (Bhargava *et al.* 2015; Ghani *et al.* 2018; Bedoya-Serna *et al.* 2018).

Research has shown that nanoemulsion formulations containing essential oil of *Mentha pulegium* (pennyroyal) (Damani *et al.* 2022), *Capsicum annum* L. (pepper) (El-Naggar *et al.* 2020), *Myrtus communis* L. (myrtle) (Roozitalab *et al.* 2022), *Allium sativum* (garlic) (Hassan; Mujtaba, 2019), *Cinnamomum* spp. (cinnamon) (Valizadeh *et al.* 2018) and *Citrus paradisi* (grapefruit) (Wang *et al.* 2021) have shown antimicrobial activity against *Staphylococcus aureus* and *Escherichia coli*. It is important to note that the authors reported that the nanoemulsions based on the essential oils of *M. pulegium*, *C. annum*, *M. communis*, *A. sativum* and *Cinnamonum* spp. showed no enhancement effect in comparison with the free form of essential oils in antibacterial tests. In contrast, the nanoemulsion based on the essential oil of *C. paradisi* had a significantly lower minimum inhibitory concentration (MIC) than the free essential oil. In this respect, many reports have shown that the conversion of spice extracts or essential oils into nanoemulsions greatly improves their antibacterial activity, presumably because the small lipid droplets interact better with microbial cells.

In this respect, it is relevant to highlight the research data of Zamaniahari *et al.* (2022), when investigating the antimicrobial effect of *Mentha spicata* (mint) against pathogens of medical importance. The researchers found that the MIC of the free essential oil was greater than that of the nanoemulsion against *S. typhimurium*, with concentrations of 6 mg/mL of the essential oil in free form and 2 mg/mL in nanoemulsion form. They also reported similar results for *S. aureus* (5mg/mL vs. 2mg/mL), *E. coli* (6mg/mL vs. 2mg/mL), *B. cereus* (4mg/mL vs. 2mg/mL) and *Listeria monocytogenes* (5mg/mL vs. 1mg/mL), respectively. The authors further stated that the bacterial cells were greatly disintegrated and that levels of cell membrane damage were caused by the nanoencapsulated oil against all bacterial species in this study.

Wei *et al.* (2022) analyzed the antimicrobial effects of nanoemulsions carried with the essential oil of *Melaleuca alternifolia* (melaleuca) and conjugated with antibiotics against six clinical strains of multiresistant *E. coli*. They observed, curiously, that the antibacterial activity of unblended nanoemulsion showed similar results for all multidrug resistant strains of *E. coli* and the reference strain (*E. coli* ATCC 25922) with MIC values of 1 mg/mL. The results of the synergism assays of the nanoemulsion combined with the standard drug had synergy with several antibiotics against multidrug-resistant *E. coli* strains. The researchers also stated that the nanoemulsion combined with doxycycline exhibited bactericidal activity, and that usually bactericidal agents can be clinically recommended instead of bacteriostatic agents, since they can reduce the development of resistance and the spread of infection.

Nanoemulsions prepared with the essential oil of *Artemisia annua* (chamomile), using Tween 80 as surfactant agent, had particles with an average diameter of 160 nm. These were tested to characterize their antimicrobial activity in gram-positive and gram-negative bacteria, as well as in *Candida* species. The nanoemulsions exhibited notable antibacterial and antifungal activities against various microbial strains, with MICs ranging from $1.42 \pm 0.64 \mu\text{g/mL}$ to $3.15 \pm 0.16 \mu\text{g/mL}$ against bacterial strains and from $3.62 \pm 0.65 \mu\text{g/mL}$ to $4.29 \pm 0.82 \mu\text{g/mL}$ against yeasts (Das *et al.*, 2020). They also observed antimicrobial effects of nanoemulsions based on essential oil of *Citrus limonum* (lemon), Tween 80 and water,

which generated nanoemulsions with a mean particle diameter of 181.5 nm. These had minimum inhibitory concentrations ranging from 3,125 to 25 mg/mL against the bacteria *S. aureus*, *Klebsiella pneumoniae*, *Salmonella typhimurium*, *Enterococcus faecalis*, *Photobacterium damsela*, *Vibrio vulnificus*, *Proteus mirabilis*, *Serratia liquefacien* and *Pseudomonas luteola* (Yazgan *et al.* 2019).

Some authors have suggested that the size of the droplets forming the nanoemulsion can affect the desired biological effect, among them the antimicrobial activity, and that this effect depends not only on the individual chemical components used in the formulations, but also on the system's structure (Baker *et al.* 2003).

Marinkovic *et al.* (2022), investigating the activity against *E. faecalis* of nanoemulsions composed of the essential oil of *C. martinii* (palmarosa) in different concentrations, observed that the most and least efficient nanoemulsions were those with concentrations of 6 mg/mL and 2.5 mg/mL, which had respective droplet sizes of 87 nm and MIC of 0.37 mg/mL versus droplets of 12 nm in diameter and MIC of 2.45 mg/mL. Thus, they noted that the size of the particles was linked to the amount of essential oil used in the formulations, and the antimicrobial effectiveness was not always linked to smaller particle size.

Nanoemulsions with antifungal activity have also been analyzed. Silva *et al.* (2020) evaluated the antifungal activity of nanoemulsions of the essential oil of *Copaifera langsdorffii* (copaíba), using Tween 80 and water, resulting in nanoparticles with an average diameter ranging between 641.3 and 29.98 nm, which showed inhibitory activity against the species *Paracoccidioides lutzii*, *P. brasiliensis*, *P. americana* and *P. restrepiensis*. The nanoemulsion presented MIC value of 125 µg/mL for all studied species. The encapsulation of *Coriandrum sativum* (coriander) essential oil in a chitosan nano-matrix, with an encapsulated nanoparticle size of 57–80 nm, was studied against the fungal species *Aspergillus spp.*, *Alternaria alternata*, *Penicillium spp.*, *Mycelia sterilia*, *Clasoporium herbarum* and *Fusarium spp.*, and the oil-encapsulated nanoemulsion showed antifungal efficacy by hampering the biosynthesis of ergosterol, the main component of the fungal cell membrane, by providing fluidity and permeability to the membrane (Das *et al.* 2019).

The antifungal activity of nanoemulsions containing the essential oil of *Croton cajucara* (sacaca), using pluronic as surfactant, generating droplets with an average size of 40 nm, was evaluated against the fungal species *Absidia cylindrospora*, *Cunninghamella elegans*, *Mucor* spp, *Rhizopus* spp, *Syncephalastrum racemosum* and *C. albicans*, with minimum inhibitory concentrations ranging from 12.21 to 195.31 µg/mL against the studied fungal species (Azevedo *et al.* 2021).

The essential oil of *Lippia sidoides* (alecrim pepper) was loaded into lipid-based systems, such as oleic acid along with Compritol and sodium dodecyl sulfate. The authors determined a particle size range of 213.1-445.5 nm. Lipid nanoparticles loaded with the essential oil of *L. sidoides* showed minimum inhibitory concentrations between 281 and 563 µg/mL against multiresistant *C. auris* (Baldim *et al.* 2022).

The potential has been studied of nanoemulsions synthesized from the ethanolic extract of *Cuphea ignea* (flor-de-santo-antonio), in addition to formulations with the essential oil of *Jasminum humile* L. (jasmine) and ethanolic extract of *Ginkgo biloba*, with findings of antiviral activity against important viruses that cause human infections: SARS CoV-2 (Covid-19) (Mahmoud *et al.* 2021), HAV (hepatitis A virus), HSV (herpes virus) (Mansour *et al.* 2022), H3N2 (influenza virus) and HBV (hepatitis B virus) (Wang *et al.* 2015).

The investigation of physicochemical properties and the carrying capacity of nanoparticle emulsions has led to successful antiviral applications administered by different routes in several *in vitro* and *in vivo* studies. A recent *in vitro* study showed that the cationic particles of a nanoemulsion loaded with *Curcumin* spp. (golden ginger) increased the antiviral activity of *Curcumin* spp. against dengue virus serotypes and improved biocompatibility with target cells (Nabila *et al.* 2020). The cationic surfaces of the particles facilitated binding to negatively charged infected and uninfected cells, improving load delivery and therapeutic efficiency.

Traditional antiviral drugs are limited by their low solubility, rapid clearance profiles, low bioavailability, nonspecific targeting, and high toxicity (Shah, 2021). Nanoemulsions can overcome these limitations because these systems are adjustable, with versatile oil cores,

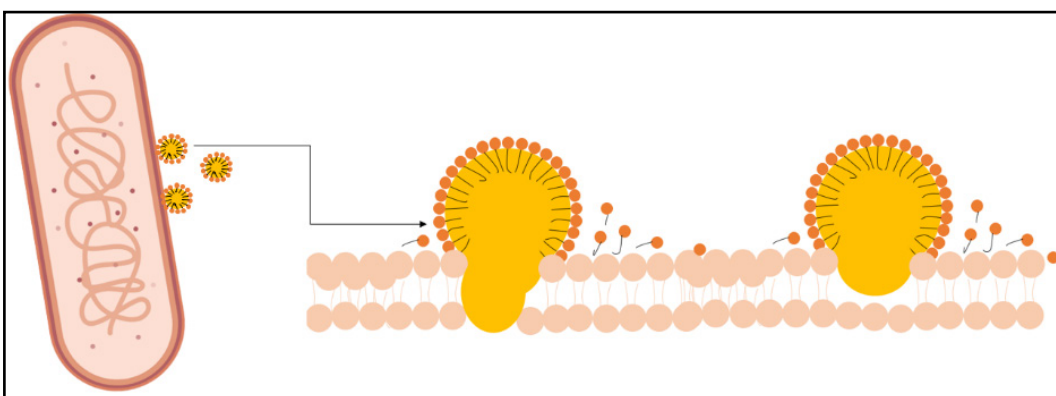
improved encapsulation capacity, easy surface modification and flexible dosage forms (Delshadi *et al.* 2021).

2.4 Nanoemulsions as carriers of natural products and their mechanism of antimicrobial action

Most natural products have low solubility, posing one of the biggest obstacles to the development of usable drugs, such as antimicrobials. This causes extremely low bioavailability, high dosage level and low selectivity (Nielsen *et al.* 2004). For an antimicrobial compound to be effective, it must interact with the specific target and have minimal interactions with unwanted targets, to increase its potency and reduce adverse effects.

Several studies have sought to avoid these effects by encapsulating antimicrobials in colloidal delivery systems, such as nanoemulsions (Weiss *et al.* 2009; Acevedo-Fani *et al.* 2017b; Khaneghah *et al.* 2018). Encapsulation involves trapping substances within small micelles to enable their antimicrobial effect (El-Kader; Abu Hashish, 2020). Figure 4 represents the interaction of nanoparticles of a nanoemulsion with the microbial cell membrane through simple diffusion.

Figure 4 – Scheme of delivery of nanoemulsion nanoparticles to the microbial cell membrane



Source: Authors' private collection (February 2023)

The literature suggests that the internalization of substances present in nanoparticles into microbial cells occurs through simple diffusion, a process that is

caused by the passive movement of nanoparticles from a more concentrated place to a less concentrated one, without energy expenditure (Harun *et al.* 2018; To *et al.* 2023).

Their use as drug carriers is promising because it allows the controlled release of an active principle at a specific place in the body for a longer period of time than conventional administration, maintaining the proper therapeutic concentration and avoiding the need for repeated administration (Park *et al.* 1999; Kumar, 2000; Kshisagar *et al.* 2005). Figure 5 shows a comparison of plasma drug levels achieved in conventional therapeutic administration and via controlled release administration.

The benefits include increased selectivity and reduced side effects (Kshisagar, 2005; Wang *et al.* 2008; Bedin *et al.* 2011), increased apparent solubility, biocompatibility and protection against physical and chemical degradation of the active principle, inducing increased pharmacological action and protection against toxicity, increased dispersibility in water, resistance to environmental conditions, and antimicrobial potency (Ray *et al.* 2016; Ferreira; Nunes, 2019).

The hydrophobicity of dispersants and surfactants together with the phytochemicals present in essential oils collectively contribute to the antimicrobial activities of nanoemulsions. These activities occur due to action on one or more binding targets in the microbial cell. Knowing the target of action is of fundamental importance to ensure that nanoemulsions have specificity for pathogens and that they will not bind to host cells causing unwanted effects.

Currently known antimicrobials act mainly on membrane and cell wall targets. In bacteria, the targets are inhibition of cell wall precursors, binding to ribosomes to prevent synthesis of essential proteins, inhibition of nucleic acid synthesis, inhibition of the metabolic pathway, and membrane disruption caused by inhibition of important cell membrane precursors (Purssel, 2019). In fungi, the most common mechanisms include inhibition of ergosterol biosynthesis (azole derivatives), inhibition of DNA replication and protein synthesis (5-fluorocytosine), binding to ergosterol present in the membrane creating pores that culminate in cell leakage (polyenes) and inhibition

of the synthesis of 1,3- β -D-glucan, which constitutes the cell wall (echinocandins) (Ivanov *et al.* 2022).

Antifungal drugs usually have important cytotoxic effects due to the similarity of fungal and human cells (both are eukaryotic). Therefore, the search for compounds that have specific targets of action in fungal cells is a challenge.

Krishnamoorthy *et al.* (2021) presented evidence that a nanoemulsion inhibited the synthesis by *C. albicans* of chitin, an important polysaccharide that makes up the yeast wall. Microscopic images showed disruption of the yeast cell wall, probably caused by inhibition of cell wall polysaccharides. In turn, Pannu *et al.* (2009) reported that NB-002, a nanoemulsion with activity against filamentous fungi, dermatophytes and *C. albicans*, had a fungistatic effect, and microscopic images demonstrated that the compound caused rupture of the fungal wall of *T. rubrum*.

Similar to what occurs in fungi, in bacteria nanoemulsions cause membrane disruption. Several studies using micrographs obtained from scanning electron microscopy (SEM) and protein leak tests supported this evidence (Moghimi *et al.* 2016; Yang *et al.* 2022). This rupture causes irreversible damage to the bacterial membrane, culminating in cell lysis. This mechanism was observed in both gram-negative and gram-positive bacteria.

The physicochemical properties of surfactants influence the mechanism of action of nanoemulsions. In gram-negative bacteria, some surfactants can better translocate through the peptidoglycan outer membrane and accumulate in the inner membrane (Sharma *et al.* 2021). Cationic surfactants have better activity compared to nanoemulsions with anionic surfactants because this property provides an electrostatic interaction with the negative membrane of bacteria (Al-Adham *et al.* 2021).

3 CONCLUSIONS

Nanoemulsions loaded with essential oils and plant extracts from the different botanical species reported here showed *in vitro* antimicrobial activity against different

microorganisms of medical importance, in addition to enhancing the antimicrobial effects of these bioproducts. Therefore, nanostructured antimicrobials with essential oils and plant extracts can be considered treatment options for microbial diseases. Due to their physicochemical properties, they will provide better delivery of natural products with good bioavailability, by reducing their toxicity and prolonging the useful life of natural antimicrobials, improving treatment of infectious human diseases.

However, more studies are needed regarding the methods and conditions for production of nanoemulsions, as well as their properties and safety, because of the limited understanding of their mechanisms and consequences on health. The consideration of all these factors is necessary for the production of nanoemulsions on an industrial scale, in turn for the development of products with better quality and safety and lower costs.

CONFLICTS OF INTEREST

The authors declare no conflicts of interest.

REFERENCES

ABREU, F. O. M. D. S.; COSTA, E. F.; CARDIAL, M. R. L.; ANDRÉ W. P. P. Polymeric nanoemulsions enriched with *Eucalyptus citriodora* essential oil. *Polímeros*, 30, 2022. <https://doi.org/10.1590/0104-1428.00920>

ACEVEDO-FANI, A.; SOLIVA-FORTUNY, R.; MARTÍN-BELLOSO, O. Nanostructured emulsions and nanolaminates for delivery of active ingredients: Improving food safety and functionality. *Trends in Food Science & Technology*, 60, 12-22, 2017. <https://doi.org/10.1016/j.tifs.2016.10.027>

AL-ADHAM, I. S.; JABER, N.; AL-REMAWI, M.; AL-AKAYLEH, F.; AL-KAISSI, E.; ALI AGHA, A. S. A.; ... & COLLIER, P. J. A review of the antimicrobial activity of thermodynamically stable microemulsions. *Letters in Applied Microbiology*, 75(3), 537-547, 2022. <https://doi.org/10.1111/lam.13570>

ANTON, N.; BENOIT, J. P.; SAULNIER, P. Design and production of nanoparticles formulated from nano-emulsion templates—a review. *Journal of controlled release*, 128(3), 185-199, 2008. <https://doi.org/10.1016/j.jconrel.2008.02.007>

AZEVEDO, M. M.; ALMEIDA, C. A.; CHAVES, F. C.; RICCI-JÚNIOR, E.; GARCIA, A. R.; RODRIGUES, I. A.; ... ALVIANO, D. S. *Croton cajucara* essential oil nanoemulsion and its antifungal activities. *Processes*, 9(11), 2021. <https://doi.org/10.3390/pr9111872>

BAKER, J. R.; HAMOUDA, T.; SHIH, A.; MYC, A. US Patent No. 6,559,189, 2003.

BALDIM, I.; PAZIANI, M. H.; GRIZANTE-BARIÃO, P. H.; KRESS, M. R. V. Z.; OLIVEIRA, W. P. Nanostructured lipid carriers loaded with *Lippia sidoides* essential oil as a strategy to combat the multidrug-resistant *Candida auris*. *Pharmaceutics*, 14(1), 180, 2022. <https://doi.org/10.3390/pharmaceutics14010180>

BAZANA, M. T.; CODEVILLA, C. F.; MENEZES, C. R; Nanoencapsulation of bioactive compounds: Challenges and perspectives. **Current opinion in food science**, 26, 47-56, 2019. <https://doi.org/10.1016/j.cofs.2019.03.005>

BEDIN, A. C. Nanoemulsões contendo benzoilmetronidazol: desenvolvimento, caracterização e estudo de liberação *in vitro*, 2011.

BEDOYA-SERNA, C. M.; DACANAL, G. C.; FERNANDES, A. M.; PINHO, S. C. Antifungal activity of nanoemulsions encapsulating oregano (*Origanum vulgare*) essential oil: *in vitro* study and application in Minas Padrão cheese. **Brazilian journal of microbiology**, 49, 929-935, 2018. <https://doi.org/10.1016/j.bjm.2018.05.004>

BHARGAVA, K.; CONTI, D. S.; ROCHA, S. R.; ZHANG, Y. Application of an oregano oil nanoemulsion to the control of foodborne bacteria on fresh lettuce. **Food microbiology**, 47, 69-73, 2015. <https://doi.org/10.1016/j.fm.2014.11.007>

BOIRE, N. A.; RIEDEL, S.; PARRISH, N. M. Essential oils and future antibiotics: new weapons against emerging 'superbugs'. **J Anc Dis Prev Rem**, 1(2), 105, 2013. <http://dx.doi.org/10.4172/jadpr.1000105>

BOTELHO, B. O.; MELO, D. C. A.; FONTES, C.; QUEIROZ, V. T.; COSTA, A. V.; MARTINS, I. V. F. Aplicação de nanoemulsões na agricultura e medicina veterinária. **TÓPICOS ESPECIAIS EM CIÊNCIA ANIMAL VII**, 143, 2018.

CHINNAIYAN, S. K.; PANDIYAN, R.; NATESAN, S.; CHINDAM, S.; GOUTI, A. K.; SUGUMARAN, A. Fabrication of basil oil Nanoemulsion loaded gellan gum hydrogel—Evaluation of its antibacterial and anti-biofilm potential. **Journal of Drug Delivery Science and Technology**, 68, 103129, 2022. <https://doi.org/10.1016/j.jddst.2022.103129>

D'AGOSTINO, M.; TESSE, N.; FRIPPIAT, J. P.; MACHOUART, M.; DEBOURGOGNE, A. Essential oils and their natural active compounds presenting antifungal properties. **Molecules**, 24(20), 3713, 2019. <https://doi.org/10.3390/molecules24203713>

DAMANI, M. H.; PARTOVI, R.; SHAHAVI, M. H.; AZIZKHANI, M. Nanoemulsions of *Trachyspermum copticum*, *Mentha pulegium* and *Satureja hortensis* essential oils: formulation, physicochemical properties, antimicrobial and antioxidant efficiency. **Journal of Food Measurement and Characterization**, 16(3), 1807-1819, 2020. <https://doi.org/10.1007/s11694-022-01294-5>

DANIELLI, L. J.; REIS, M.; BIANCHINI, M.; CAMARGO, G. S.; BORDIGNON, S. A.; GUERREIRO, I. K.; ... APEL, M. A. Antidermatophytic activity of volatile oil and nanoemulsion of *Stenachaenium megapotamicum* (Spreng.) Baker. **Industrial crops and products**, 50, 23-28, 2013. <https://doi.org/10.1016/j.indcrop.2013.07.027>

DAS, S.; HORVÁTH, B.; ŠAFRANKO, S.; JOKIĆ, S.; SZÉCHENYI, A.; KÓSZEGI, T. Antimicrobial activity of chamomile essential oil: Effect of different formulations. **Molecules**, 24(23), 4321, 2019. <https://doi.org/10.3390/molecules24234321>

DAS, S.; SINGH, V. K.; DWIVEDY, A. K.; CHAUDHARI, A. K.; UPADHYAY, N.; SINGH, P.; ... DUBEY, N. K. Encapsulation in chitosan-based nanomatrix as an efficient green technology to boost the antimicrobial, antioxidant and in situ efficacy of *Coriandrum sativum* essential oil. **International journal of biological macromolecules**, 133, 294-305, 2019. <https://doi.org/10.1016/j.ijbiomac.2019.04.070>

DAS, S.; VÖRÖS-HORVÁTH., B; BENCSIK., T; MICALIZZI, G.; MONDELLO, L.; HORVÁTH, G.; ... SZÉCHENYI, A. Antimicrobial activity of different Artemisia essential oil formulations. **Molecules**, 25(10), 2390, 2020. <https://doi.org/10.3390/molecules25102390>

DELSHADI, R.; BAHRAMI, A.; MCCLEMENTS, D. J.; MOORE, M. D.; WILLIAMS, L. Development of nanoparticle-delivery systems for antiviral agents: A review. **Journal of Controlled Release**, 331, 30-44, 2021. <https://doi.org/10.1016/j.jconrel.2021.01.017>

DONSÌ, F; FERRARI, G. Essential oil nanoemulsions as antimicrobial agents in food. **Journal of biotechnology**, 233, 106-120, 2016. <https://doi.org/10.1016/j.jbiotec.2016.07.005>

EL-KADER, A.; ABU HASHISH, H. Encapsulation techniques of food bioproduct. **Egyptian Journal of Chemistry**, 63(5), 1881-1909, 2020. <https://doi.org/10.21608/ejchem.2019.16269.1993>

EL-NAGGAR, M. E.; SOLIMAN, R. A.; MORSY, O. M.; ABDEL-AZIZ, M. S. Nanoemulsion of Capsicum fruit extract as an eco-friendly antimicrobial agent for production of medical bandages. **Biocatalysis and Agricultural Biotechnology**, 23, 101516, 2020. <https://doi.org/10.1016/j.bcab.2020.101516>

FERREIRA, C. D.; NUNES, I. L. Oil nanoencapsulation: development, application, and incorporation into the food market. **Nanoscale research letters**, 14, 1-13., 2019. <https://doi.org/10.1186/s11671-018-2829-2>

FRANZOL, A.; REZENDE, M. C. Estabilidade de emulsões: um estudo de caso envolvendo emulsionantes aniônico, catiônico e não-iônico. **Polímeros**, 25, 1-9, 2015. <https://doi.org/10.1590/0104-1428.1669>

GALIE, S.; GARCÍA-GUTIÉRREZ, C.; MIGUÉLEZ, E. M.; VILLAR, C. J.; LOMBÓ, F. Biofilms in the food industry: health aspects and control methods. **Frontiers in microbiology**, 9, 898, 2018. <https://doi.org/10.3389/fmicb.2018.00898>

GHANI, S.; BARZEGAR, H.; NOSHAD, M.; HOJJATI, M. The preparation, characterization and in vitro application evaluation of soluble soybean polysaccharide films incorporated with cinnamon essential oil nanoemulsions. **International journal of biological macromolecules**, 112, 197-202, 2018. <https://doi.org/10.1016/j.ijbiomac.2018.01.145>

GHAZY, O. A.; FOUAD, M. T.; SALEH, H. H.; KHOLIF, A. E.; MORSY, T. A. Ultrasound-assisted preparation of anise extract nanoemulsion and its bioactivity against different pathogenic bacteria. **Food chemistry**, 341, 128259, 2021. <https://doi.org/10.1016/j.foodchem.2020.128259>

GUERRA-ROSAS, M. I.; MORALES-CASTRO, J.; OCHOA-MARTÍNEZ, L. A.; SALVIA-TRUJILLO, L.; MARTÍN-BELLOSO, O. Long-term stability of food-grade nanoemulsions from high methoxyl pectin containing essential oils. **Food Hydrocolloids**, 52, 438-446, 2016. <https://doi.org/10.1016/j.foodhyd.2015.07.017>

HARUN, S. N.; NORDIN, S. A.; ABD GANI, S. S.; SHAMSUDDIN, A. F.; BASRI, M.; BASRI, H. B. Development of nanoemulsion for efficient brain parenteral delivery of cefuroxime: Designs, characterizations, and pharmacokinetics. **International journal of nanomedicine**, 13, 2571, 2018. <https://doi.org/10.2147/IJN.S151788>

HASSAN, K. A.; MUJTABA, M. A. Antibacterial efficacy of garlic oil nano-emulsion. **AIMS Agriculture and Food**, 4(1), 194-205, 2019. <https://doi.org/10.3934/agrfood.2019.1.194>

HERCULANO, E. D.; PAULA, H. C.; FIGUEIREDO, E. A.; DIAS, F. G.; PEREIRA, V. D. A. Physicochemical and antimicrobial properties of nanoencapsulated *Eucalyptus staigeriana* essential oil. **LWT-Food Science and Technology**, 61(2), 484-491, 2015. <https://doi.org/10.1016/j.lwt.2014.12.001>

ISAH, T. Stress and defense responses in plant secondary metabolites production. **Biological research**, 52, 2019. <https://dx.doi.org/10.1186/s40659-019-0246-3>

IVANOV, M.; ĆIRIĆ, A.; STOJKOVIĆ, D. Emerging antifungal targets and strategies. **International Journal of Molecular Sciences**, 23(5), 2756, 2022. <https://doi.org/10.3390/ijms23052756>

JEONG, S. H.; HUH, K. M.; PARK, K. Hydrogel drug delivery systems. In **Polymers in drug delivery** (pp. 49-62), 2006.

JEROBIN, J.; MAKWANA, P.; SURESH KUMAR, R. S.; SUNDARAMOORTHY, R.; MUKHERJEE, A.; CHANDRASEKARAN, N. Antibacterial activity of neem nanoemulsion and its toxicity assessment on human lymphocytes *in vitro*. **International journal of nanomedicine**, 10(sup2), 77-86, 2015.

KALEELULLAH, R. A.; GARUGULA, N. Teratogenic Genesis in Fetal Malformations. **Cureus**, 13(2), 2021. <https://doi.org/10.7759/cureus.13149>

KALOYANIDES, G. J. Antibiotic-related nephrotoxicity. *Nephrology, Dialysis, Transplantation: Official Publication of the European Dialysis and Transplant Association-European Renal Association*, 9, 130-134, 1994.

KARIM, A.; REHMAN, A.; FENG, J.; NOREEN, A.; ASSADPOUR, E.; KHARAZMI, M. S.; JAFARI, S. M. Alginate-based nanocarriers for the delivery and controlled-release of bioactive compounds. **Advances in Colloid and Interface Science**, 102744, 2022. <https://doi.org/10.1016/j.cis.2022.102744>

KEMBER, M.; GRANDY, S.; RAUDONIS, R.; CHENG, Z. Non-canonical host intracellular niche links to new antimicrobial resistance mechanism. **Pathogens**, 11(2), 220, 2022. <https://doi.org/10.3390/pathogens11020220>

KHANEGHAH, A. M.; HASHEMI, S. M. B.; LIMBO, S. Antimicrobial agents and packaging systems in antimicrobial active food packaging: An overview of approaches and interactions. **Food and Bioproducts Processing**, 111, 1-19, 2018. <https://doi.org/10.1016/j.fbp.2018.05.001>

KRISHNAMOORTHY, R.; GASSEM, M. A.; ATHINARAYANAN, J.; PERIYASAMY, V. S.; PRASAD, S.; ALSHATWI, A. A. Antifungal activity of nanoemulsion from *Cleome viscosa* essential oil against food-borne pathogenic *Candida albicans*. **Saudi Journal of Biological Sciences**, 28(1), 286-293, 2021. <https://doi.org/10.1016/j.sjbs.2020.10.001>

KSHIRSAGAR, N. A.; PANDYA, S. K.; KIRODIAN, B. G.; SANATH, S. Liposomal drug delivery system from laboratory to clinic. **Journal of postgraduate medicine**, 51(5), 5, 2015.

KUMAR, N.; MANDAL, A. Surfactant stabilized oil-in-water nanoemulsion: stability, interfacial tension, and rheology study for enhanced oil recovery application. **Energy & fuels**, 32(6), 6452-6466, 2018. <https://doi.org/10.1021/acs.energyfuels.8b00043>

KUMAR, M. N. R. A review of chitin and chitosan applications. **Reactive and functional polymers**, 46(1), 1-27, 2000. [https://doi.org/10.1016/S1381-5148\(00\)00038-9](https://doi.org/10.1016/S1381-5148(00)00038-9)

LEÃO, K. M. M.; REIS, L. V. C.; SPERANZA, P.; RODRIGUES, A. P.; RIBEIRO, A. P. B.; MACEDO, J. A.; MACEDO, G. A. Physicochemical characterization and antimicrobial activity in novel systems containing buriti oil and structured lipids nanoemulsions. **Biotechnology reports**, 24, e00365, 2019. <https://doi.org/10.1016/j.btre.2019.e00365>

LIMA, T. P.; SOUSA, T. L.; OLIVEIRA, J. P. M.; FELIZARDO, M. G. A. C.; EVERTON, G. O.; MOUCHREK FILHO, V. E. Chemical profile, thermodynamic stability and fungicidal activity of the nanoemulsion incorporated with essential oil and hydroalcoholic extract of *Syzygium aromaticum* (L.) Merr. & LM. Perry. **Ciência e Natura**, 43, 77, 2021. <https://doi.org/10.5902/2179460X63929>

MACCELLI, A.; VITANZA, L.; IMBRIANO, A.; FRASCHETTI, C.; FILIPPI, A.; GOLDONI, P.; ... RINALDI, F. *Satureja montana* L. Essential oils: Chemical profiles/phytochemical screening, antimicrobial activity and o/w nanoemulsion formulations. **Pharmaceutics**, 12(1), 7, 2019. <https://doi.org/10.3390/pharmaceutics12010007>

MADIGAN, M. T.; MARTINKO, J. M.; BENDER, K. S.; BUCKLEY, D. H.; STAHL, D. A. **Brock's Microbiology-14^a Edition**. Artmed Publisher, 2016.

MAHMOUD, D. B.; ISMAIL, W. M.; MOATASIM, Y.; KUTKAT, O.; ELMESHAD, A. N.; EZZAT, S. M.; ... MOSTAFA, A. Delineating a potent antiviral activity of *Cuphea ignea* extract loaded nano-formulation against SARS-CoV-2: In silico and *in vitro* studies. **Journal of Drug Delivery Science and Technology**, 66, 102845, 2021. <https://doi.org/10.1016/j.jddst.2021.102845>

MANSOUR, K. A.; EL-NEKETI, M; LAHLOUB, M. F.; ELBERMAWI, A. Nanoemulsions of *Jasminum humile* L. and *Jasminum grandiflorum* L. Essential Oils: An Approach to Enhance Their Cytotoxic and Antiviral Effects. **Molecules**, 27(11), 3639, 2022. <https://doi.org/10.3390/molecules27113639>

MAPHOSA, Y.; JIDEANI, V. A. Factors affecting the stability of emulsions stabilised by biopolymers. **Science and technology behind Nanoemulsions**, 65, 2018. <https://doi.org/10.5772/intechopen.75308>

MARINKOVIĆ, J.; BOŠKOVIĆ, M.; TASIĆ, G.; VASILJEVIĆ, B.; MARKOVIĆ, D.; MARKOVIĆ, T.; NIKOLIĆ, B. *Cymbopogon martinii* essential oil nanoemulsions: Physico-chemical characterization, antibacterial and antibiofilm potential against *Enterococcus faecalis*. **Industrial Crops and Products**, 187, 115478, 2022. <https://doi.org/10.1016/j.indcrop.2022.115478>

MOAZENI, M.; DAVARI, A.; SHABANZADEH, S.; AKHTARI, J.; SAEEDI, M.; MORTYEZA-SEMNANI, K.; ... NOKHODCHI, A. *In vitro* antifungal activity of *Thymus vulgaris* essential oil nanoemulsion. **Journal of Herbal Medicine**, 28, 100452, 2021. <https://doi.org/10.1016/j.hermed.2021.100452>

MOGHIMI, R.; ALIAHMADI, A.; MCCLEMENTS, D. J.; RAFATI, H. Investigations of the effectiveness of nanoemulsions from sage oil as antibacterial agents on some food borne pathogens. **LWT-Food Science and Technology**, 71, 69-76, 2016. <https://doi.org/10.1016/j.lwt.2016.03.018>

MOSTAFA, A. A.; AL-ASKAR, A. A.; ALMAARY, K. S.; DAWOUD, T. M.; SHOLKAMY, E. N.; BAKRI, M. M. Antimicrobial activity of some plant extracts against bacterial strains causing food poisoning diseases. **Saudi journal of biological sciences**, 25(2), 361-366, 2018. <https://doi.org/10.1016/j.sjbs.2017.02.004>

NABILA, N.; SUADA, N. K.; DENIS, D.; YOHAN, B.; ADI, A. C.; VETERINI, A. S.; ... RACHMAWATI, H. Antiviral action of curcumin encapsulated in nanoemulsion against four serotypes of dengue virus. **Pharmaceutical Nanotechnology**, 8(1), 54-62, 2020. <https://doi.org/10.2174/2211738507666191210163408>

NASCIMENTO, J. F.; COSTA, E. F.; ABREU, F. O. M. S. CARACTERIZAÇÕES DE NANOEMULSÕES DE ALGINATO DE SÓDIO COM ÓLEO ESSENCIAL DE *Eucalyptus citriodora*. **Revista Coleta Científica**, 4(8), 15-22, 2020. <https://doi.org/10.7910/DVN/7QGVTR>

NIELSEN, S. F.; BOESEN, T.; LARSEN, M.; SCHØNNING, K.; KROMANN, H. Antibacterial chalcones—bioisosteric replacement of the 4'-hydroxy group. **Bioorganic & medicinal chemistry**, 12(11), 3047-3054, 2004. <https://doi.org/10.1016/j.bmc.2004.03.071>

OKONKWO, S.; EMEJE, M.; PETERS, O.; OKHALE, S. Extraction and Nanoencapsulation of *Ocimum Gratissimum* Leaf Extract and Its Anti-Mycobacterial Activities. **Extraction**, 12(2), 2020. <https://doi.org/10.7176/CMR/12-2-03>

OUN, R.; MOUSSA, Y. E.; WHEATE, N. J. The side effects of platinum-based chemotherapy drugs: a review for chemists. **Dalton transactions**, 47(19), 6645-6653, 2018. <https://doi.org/10.1039/C8DT90088D>

PAHO - **Pan American Health Organization**. WHO reveals the main causes of death and disability worldwide between 2000 and 2019. December 9, 2020. Available at: < <https://www.paho.org/pt/noticias/9-12-2020-oms-reveals-main-causes-of-death-and-disability-and-m-all-world-between-2000-and>>. Accessed on: November 13, 2021

PAIM, L. F. N. A.; DALLA LANA, D. F.; GIARETTA, M.; DANIELLI, L. J.; FUENTEFRIA, A. M.; APEL, M. A.; KÜLKAMP-GUERREIRO, I. C. *Poiretia latifolia* essential oil as a promising antifungal and anti-inflammatory agent: Chemical composition, biological screening, and development of a nanoemulsion formulation. **Industrial crops and products**, 126, 280-286, 2018. <https://doi.org/10.1016/j.indcrop.2018.10.016>

PANNU, J.; MCCARTHY, A.; MARTIN, A.; HAMOUDA, T.; CIOTTI, S.; FOTHERGILL, A.; SUTCLIFFE, J. NB-002, a novel nanoemulsion with broad antifungal activity against dermatophytes, other filamentous fungi, and *Candida albicans*. **Antimicrobial agents and chemotherapy**, 53(8), 3273-3279, 2019. <https://doi.org/10.1128/AAC.00218-09>

PATHANIA, R.; NAJDA A.; CHAWLA, P.; KAUSHIK, R.; KHAN, M. A. Low-energy assisted sodium alginate stabilized *Phyllanthus niruri* extract nanoemulsion: Characterization, in vitro antioxidant and antimicrobial application. **Biotechnology Reports**, 33, e00711, 2022. <https://doi.org/10.1016/j.btre.2022.e00711>

PURSSELL, E. Antimicrobials. *Understanding Pharmacology in Nursing Practice*, 147-165, 2019.

QUATRIN, P. M.; VERDI, C. M.; SOUZA, M. E.; GODOI, S. N.; KLEIN, B.; GUNDEL, A.; ... SANTOS, R. C. V. Antimicrobial and antibiofilm activities of nanoemulsions containing *Eucalyptus globulus* oil against *Pseudomonas aeruginosa* and *Candida* spp. **Microbial pathogenesis**, 112, 230-242, 2017. <https://doi.org/10.1016/j.micpath.2017.09.062>

QUINTÃO, F. J.; TAVARES, R. S.; VIEIRA-FILHO, S. A.; SOUZA, G. H.; SANTOS, O. D. Hydroalcoholic extracts of *Vellozia squamata*: study of its nanoemulsions for pharmaceutical or cosmetic applications. **Revista Brasileira de Farmacognosia**, 23(1), 101-107, 2013. <https://doi.org/10.1590/S0102-695X2013005000001>

RABINOVICH-GUILATT, L.; COUVREUR, P.; LAMBERT, G.; GOLDSTEIN, D.; BENITA, S.; DUBERNET, C. Extensive surface studies help to analyse zeta potential data: the case of cationic emulsions. **Chemistry and Physics of Lipids**, 131(1), 1-13, 2004. <https://doi.org/10.1016/j.chemphyslip.2004.04.003>

RAO, P. J.; KHANUM, H. A green chemistry approach for nanoencapsulation of bioactive compound–Curcumin. **LWT-Food Science and Technology**, 65, 695-702, 2016. <https://doi.org/10.1016/j.lwt.2015.08.070>

RAY, S.; RAYCHAUDHURI, U.; CHAKRABORTY, R. An overview of encapsulation of active compounds used in food products by drying technology. **Food bioscience**, 13, 76-83, 2016. <https://doi.org/10.1016/j.fbio.2015.12.009>

REDDY, S. R.; MELIK, D. H.; FOGLER, H. S. Emulsion stability—theoretical studies on simultaneous flocculation and creaming. **Journal of Colloid and Interface Science**, 82(1), 116-127, 1981. [https://doi.org/10.1016/0021-9797\(81\)90129-6](https://doi.org/10.1016/0021-9797(81)90129-6)

REYGAERT, W. C. An overview of the antimicrobial resistance mechanisms of bacteria. **AIMS microbiology**, 4(3), 482, 2018. <https://doi.org/10.3934/microbiol.2018.3.482>

ROOZITALAB, G.; YOUSEFPOOR, Y.; ABDOLLAHI, A.; SAFARI, M.; RASTI, F.; OSANLOO, M. Antioxidative, anticancer, and antibacterial activities of a nanoemulsion-based gel containing *Myrtus communis* L. essential oil. **Chemical Papers**, 76(7), 4261-4271, 2022. <https://doi.org/10.1007/s11696-022-02185-1>

SAEZ-LLORENS, X.; WONG, M. M. C.; CASTANO, E. DE SUMAN ONIX, DE MORÖS DAYSI., & DE ATENCIO IRIS Impact of an antibiotic restriction policy on hospital expenditures and bacterial susceptibilities: a lesson from a pediatric institution in a developing country. **The Pediatric infectious disease journal**, 19(3), 200-206, 2000.

SAHU, S.; KATIYAR, S. S.; KUSHWAH, V.; JAIN, S.; Active natural oil-based nanoemulsion containing tacrolimus for synergistic antipsoriatic efficacy. **Nanomedicine**, 13(16), 1985-1998, 2018. <https://doi.org/10.2217/nnm-2018-0135>

SAMBER, N.; KHAN, A.; VARMA, A.; MANZOOR, N. Synergistic anti-candidal activity and mode of action of *Mentha piperita* essential oil and its major components. **Pharmaceutical biology**, 53(10), 1496-1504, 2015. <https://doi.org/10.3109/13880209.2014.989623>

SHAH, S.; CHOUGULE, M. B.; KOTHA, A. K.; KASHIKAR, R.; GODUGU, C.; RAGHUVANSHI, R. S.; ... SRIVASTAVA, S. Nanomedicine based approaches for combating viral infections. **Journal of Controlled Release**, 338, 80-104, 2021. <https://doi.org/10.1016/j.jconrel.2021.08.011>

SHARMA, P.; VAIWALA, R.; PARTHASARATHI, S.; PATIL, N.; VERMA, A.; WASKAR, M.; ... AYAPPA, K. G. Interactions of surfactants with the bacterial cell wall and inner membrane: Revealing the link between aggregation and antimicrobial activity. **Langmuir**, 38(50), 15714-15728, 2022. <https://doi.org/10.1021/acs.langmuir.2c02520>

SIENKIEWICZ, M.; ŁYSAKOWSKA, M.; DENYS, P.; KOWALCZYK, E. The antimicrobial activity of thyme essential oil against multidrug resistant clinical bacterial strains. **Microbial drug resistance**, 18(2), 137-148, 2012. <https://doi.org/10.1089/mdr.2011.0080>

SILVA, L. C.; MIRANDA, M. A. C. M.; FREITAS, J. V.; FERREIRA, S. F. A.; LIMA, E. C. O.; OLIVEIRA, C. M. A.; ... PEREIRA, M. Antifungal activity of Copaíba resin oil in solution and nanoemulsion against *Paracoccidioides* spp. **Brazilian Journal of Microbiology**, 51, 125-134, 2020. <https://doi.org/10.1007/s42770-019-00201-3>

SPELLBERG, B.; BARTLETT, J.; WUNDERINK, R.; GILBERT, D. N. Novel approaches are needed to develop tomorrow's antibacterial therapies. **American journal of respiratory and critical care medicine**, 191(2), 135-140, 2015. <https://doi.org/10.1164/rccm.201410-1894OE>

SURASSMO, S.; MIN, S. G.; BEJRAPHA, P.; CHOI, M. J. Effects of surfactants on the physical properties of capsicum oleoresin-loaded nanocapsules formulated through the emulsion-diffusion method. **Food Research International**, 43(1), 8-17, 2010. <https://doi.org/10.1016/j.foodres.2009.07.008>

TADROS, T. Principles of emulsion stabilization with special reference to polymeric surfactants. **Journal of cosmetic science**, 57(2), 153-169, 2006.

TADROS, T. F. Correlation of viscoelastic properties of stable and flocculated suspensions with their interparticle interactions. **Advances in colloid and interface science**, 68, 97-200, 1996. [https://doi.org/10.1016/S0001-8686\(96\)90047-0](https://doi.org/10.1016/S0001-8686(96)90047-0)

TO, D.; KAKAR, A.; KALI, G.; WIBEL, R.; KNOLL, P.; MARX, F.; BERNKOP-SCHNÜRCH, A. Iminated aminoglycosides in self-emulsifying drug delivery systems: Dual approach to break down the microbial defense. **Journal of Colloid and Interface Science**, 630, 164-178, 2023. <https://doi.org/10.1016/j.jcis.2022.10.077>

TOPUZ, O. K.; ÖZVURAL, E. B.; ZHAO, Q.; HUANG, Q.; CHIKINDAS, M.; GÖLÜKÇÜ, M. Physical and antimicrobial properties of anise oil loaded nanoemulsions on the survival of foodborne pathogens. **Food chemistry**, 203, 117-123, 2016. <https://doi.org/10.1016/j.foodchem.2016.02.051>

TORTORA, G. J.; CASE, C. L.; FUNKE, B. R. **Microbiology-12^a Edition**. Artmed Publisher, 2016.

VALIZADEH, A.; SHIRZAD, M.; ESMAEILI, F.; AMANI, A. Increased antibacterial activity of cinnamon oil microemulsion in comparison with cinnamon oil bulk and nanoemulsion. **Nanomedicine Research Journal**, 3(1), 37-43, 2018.

WALLACE, R. J. Antimicrobial properties of plant secondary metabolites. **Proceedings of the nutrition society**, 63(4), 621-629, 2004. <https://doi.org/10.1079/PNS2004393>

WANG, C. Z.; LI, W. J.; TAO, R.; YE, J. Z.; ZHANG, H. Y. Antiviral activity of a nanoemulsion of polyphenols from ginkgo leaves against influenza A H3N2 and hepatitis B virus in vitro. **Molecules**, 20(3), 5137-5151, 2015. <https://doi.org/10.3390/molecules20035137>

WANG, L.; LI, X.; ZHANG, G.; DONG, J.; EASTOE, J. Oil-in-water nanoemulsions for pesticide formulations. **Journal of colloid and interface science**, 314(1), 230-235, 2007. <https://doi.org/10.1016/j.jcis.2007.04.079>

WANG, S.; LIANG, X.; ZHAO, W.; MI, X.; ZHANG, C.; ZHANG, W.; ... JIANG, Y. Preparation of nanoemulsion of grapefruit seed extract and evaluation of its antibacterial activity. **Journal of Food Processing and Preservation**, 46(1), e16197, 2022. <https://doi.org/10.1111/jfpp.16197>

WANG, J. J.; SUNG, K. C.; YEH, C. H.; FANG, J. Y. The delivery and antinociceptive effects of morphine and its ester prodrugs from lipid emulsions. **International journal of pharmaceutics**, 353(1-2), 95-104, 2008. <https://doi.org/10.1016/j.ijpharm.2007.11.013>

WEI, S.; TIAN, Q.; ZHAO, X.; LIU, X.; HUSIEN, H. M.; LIU, M.; ... LI, J. Tea Tree Oil Nanoemulsion Potentiates Antibiotics against Multidrug-Resistant *Escherichia coli*. **ACS Infectious Diseases**, 8(8), 1618-1626, 2022. <https://doi.org/10.1021/acsinfecdis.2c00223>

WEISS, J.; GAYSINSKY, S.; DAVIDSON, M.; MCCLEMENTS, J. Nanostructured encapsulation systems: food antimicrobials. In **Global issues in food science and technology**, 425-479, 2009. <https://doi.org/10.1016/B978-0-12-374124-0.00024-7>

YANG, L.; WEN, K. S.; RUAN, X.; ZHAO, Y. X.; WEI, F.; WANG, Q. Response of plant secondary metabolites to environmental factors. **Molecules**, 23(4), 762, 2018. <https://doi.org/10.3390/molecules23040762>

YANG, R.; MIAO, J.; ZHANG, Z.; WAN, C.; ZOU, L.; CHEN, C.; CHEN, J. Untargeted lipidomics reveals the antifungal mechanism of essential oils nanoemulsion against *Penicillium digitatum*. **LWT**, 168, 113909, 2022. <https://doi.org/10.1016/j.lwt.2022.113909>

YAZGAN, H.; OZOGUL, Y.; KULEY, E. Antimicrobial influence of nanoemulsified lemon essential oil and pure lemon essential oil on food-borne pathogens and fish spoilage bacteria. **International journal of food microbiology**, 306, 108266, 2019. <https://doi.org/10.1016/j.ijfoodmicro.2019.108266>

ZAMANIAHARI, S.; JAMSHIDI, A.; MOOSAVY, M. H.; KHATIBI, S. A. Preparation and evaluation of *Mentha spicata* L. essential oil nanoemulsion: physicochemical properties, antibacterial activity against foodborne pathogens and antioxidant properties. **Journal of Food Measurement and Characterization**, 16(4), 3289-3300, 2022. <https://doi.org/10.1007/s11694-022-01436-9>

ZHANG, F.; RAMACHANDRAN, G.; MOTHANA, R. A.; NOMAN, O. M.; ALOBAID, W. A.; RAJIVGANDHIG; MANOHARAN, N. Anti-bacterial activity of chitosan loaded plant essential oil against multi drug resistant *K. pneumoniae*. **Saudi Journal of Biological Sciences**, 27(12), 3449-3455, 2020. <https://doi.org/10.1016/j.sjbs.2020.09.025>

ZHANG, Z.; MCCLEMENTS, D. J. Overview of nanoemulsion properties: stability, rheology, and appearance. In **Nanoemulsions** (pp. 21-49). Academic Press, 2018. <https://doi.org/10.1016/B978-0-12-811838-2.00002-3>

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